



CONTROLLED AND SPONTANEOUS FERMENTATION IN COCONUT WATER AND STRAWBERRY: STRATEGIES FOR ENHANCING FUNCTIONAL VALUE.

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Abstract

Fermentation is among the most significant biotechnological operations that enhance the functional, nutritional, and sensory attributes of foods and beverages. Coconut and strawberry are substrates with high levels of bioactivity and are most suitable for controlled fermentation for producing functional beverages. Reproducibility of the fermentation results is challenging due to microbial diversity, differences in the composition of substrates, and environmental factors. This review indicates fermentation as an innovative biotechnological process enhancing the nutritional, functional, and sensory attributes of foods and beverages. By facilitating microbial conversion of sugars to desirable compounds, fermentation not only lengthens shelf life but also makes products rich with bioactive materials like polyphenols, organic acids, and antioxidants. Coconut and strawberry are rich in nutrients and phytochemical content and are suitable substrates for the production of functional fermented beverages. This review explores the significance of microbial selection, standardization of process, biochemical transformation, and scalability in ensuring consistent quality. Both controlled and spontaneous fermentation approaches are elaborately discussed—spontaneous fermentation provides intricate flavor development by native microbiota, and controlled fermentation guarantees reproducibility, safety, and maximized functional performance. Particular emphasis is placed on the significance of conventional and probiotic strains, like *Saccharomyces cerevisiae* and *Lactobacillus reuteri*, and how fermentation conditions—temperature (25–37 °C), pH, and fermentation time—are accountable for the quality of the product. Finally, the review highlights how fermentation can be optimized to release the full potential of coconut- and strawberry-derived beverages as next-generation functional foods.

Keywords - Controlled fermentation, Spontaneous Fermentation, Coconut Water, Strawberry, Probiotics, Nutritional Composition, Reproducibility

1. Introduction

Fermentation is said to be an ancient traditional, successful food preservation and value-addition practice followed globally in different civilizations. In the recent past, its application went beyond preservation itself and assumed vast significance for the improvement of nutritional quality and sensory acceptability of foods and beverages. Specifically, fruit fermentation, such as strawberry (*Fragaria × ananassa* Duch) and coconut (*Cocos nucifera*), into health function drinks is a new research field in food science and nutrition (Kruthi Doriya et al., 2022). Strawberries, among the most widely eaten berries in the world, are renowned for their high vitamin C, vitamin E, folate, potassium, flavonoids, phenolics, β -carotene, and other phytochemicals' contents with excellent antioxidant and anti-inflammatory properties (Zhiqiao Zhao et al., 2021; Bo Yang et al., 2024). Strawberries are also extremely perishable and hence bear high wastage at the post-harvest stage. Recent data indicate that world strawberry production amounted to 9,175.4 kilotons during 2021, with major producers such as the United States, Turkey, Mexico, and China dominating the market. This has led scholars and industry players to seek processing methods such as freezing, juicing, distillation, and fermentation to minimize wastage and extend the shelf life of products (Mario Bezerra et al., 2024). The coconut palm, *Cocos nucifera*, belongs to the family Arecaceae and yields the fruit commonly referred to as a coconut. Natural coconut water is a versatile beverage with multiple applications and also has medicinal value. In addition to carbohydrates, vitamins, minerals, amino acids, and phytohormones, it is an excellent source of electrolytes such as potassium and magnesium. It is free from fat, low in calories, and high in fiber, contributing to nutrient absorption and fluid replacement. Also, its cytokinins possess anti-thrombolytic, anti-cancer, and anti-aging activities (Polemer M. et al., 2017; Jean W.H. Yong et al., 2009). Coconut products are well known in various food and nutritional applications due to their rich nutritive metabolites and distinctive taste. Tender coconut water is a rich, refreshing beverage with minerals and other beneficial compounds (S.V. Ramesh et al., 2024). *Saccharomyces cerevisiae* D254 fermentation enhanced matured coconut water quality (Guanfei Zhang et al., 2018 et. al., 2017). Through this review, the applicability and efficacy of controlled fermentation and replicability in processing coconut and strawberry foods, particularly in the production of functional and fermented beverages, will be investigated. This review is targeted towards the comprehension of how certain fermentation parameters, such as microbial strain selection, temperature, pH, and Oxygen control, can be optimized to enhance product quality, safety, and consistency.

2. Nutritional Composition of Coconut and Strawberry

Table No:1 Comparative Nutrient Composition of Strawberries and Coconut Water (per 100 g)

Nutrient	Strawberry	Coconut Water	References
Organic acids			
Pyruvic acid (mg)	< 40	2.0	(Mario Bezerra et al., 2024; Beatriz Patricio Rocha et al., 2024)
Malic acid (mg)	198	0.02	(Miguel Ribeiro et al., 2024; Beatriz Patricio Rocha et al., 2024)
Citric acid (mg)	700	0.08	(Dong – Sheng Gao et al., 2016; Beatriz Patricio Rocha et al., 2024)
Minerals			
Iron (mg)	0.41	0.42	(Mario Bezerra et al., 2024; Yufeng Zhang et al., 2024)
Zinc (mg)	0.14	0.09	(Mario Bezerra et al., 2024; Yufeng Zhang et al., 2024)
Calcium (mg)	34.94	279.93	(Hasnaa Sadik et al., 2023; Beatriz Patricio Rocha et al., 2024)

Sodium (mg)	1	19.57	(Mario Bezerra et al., 2024; Beatriz Patricio Rocha et al., 2024)
Potassium (mg)	153	1,932.57	(Mario Bezerra et al., 2024; Beatriz Patricio Rocha et al., 2024)
Phosphorus (mg)	24	11.17	(Hasnaa Sadik et al., 2023; Beatriz Patricio Rocha et al., 2024)
Magnesium (mg)	13	85.13	(Hasnaa Sadik et al., 2023; Beatriz Patricio Rocha et al., 2024)

This table offers a comparative overview of major organic acids and mineral composition in strawberries and coconut water. It indicates that strawberries are much higher in organic acids like malic and citric acid, whereas coconut water is much higher in vital electrolytes and minerals, especially potassium, calcium, and magnesium. This difference emphasizes the complementary nutritional profiles of both, with strawberries providing antioxidant potential and coconut water being an excellent natural rehydration drink.

3. Co-Fermentation of Strawberry and Coconut Substrates

Fermentation is a critical metabolic process that occurs in the absence of oxygen (O₂). In fermentation, sugar is broken down in the absence of oxygen. Organic acids, gases, or alcohol are by-products formed as a consequence of fermentation (Arghya Mani, 2018). Blending coconut and strawberry while fermenting may produce beverages that have the hydrating properties of coconut water and the intense flavor and antioxidants of strawberries. Such combinations are attractive to a broader consumer segment seeking both health and sensory benefits (Carolina Saori Ishii Mauro et al., 2021). The blend of coconut and strawberry in fermented beverages is in accordance with existing market trends toward natural, functional, and probiotic beverages. The positive consumer response to such combinations indicates a promising direction for product development (Maria Thereza Carlos Fernandes et al., 2021).

4. Impact of Fermentation on Shelf Life, Flavor, and Functional Value Enhancement

Fermentation significantly improves the shelf life, taste, and functional properties of several food products (Sumeyye Saritas et al., 2024). Fermentation inhibits spoilage and pathogenic microorganisms by forming organic acids, bacteriocins, and other antimicrobial compounds. *Lactobacillus plantarum*, for instance, is said to enhance the safety and shelf life of fermented foods by producing such inhibitory compounds (Sudhanshu S Behera et al., 2018). Microbial fermentation leads to the production of Flavors such as esters, alcohols, and ketones. In fermented fruit juices, *L. plantarum* and *L. acidophilus* enhance flavor attributes through intensified volatile compounds, increased sensory attributes. (Yu-Han Yuan et al., 2024). Fermentation enhances the nutritional quality of food by boosting bioactive compounds. For example, fermentation enhances antioxidant activity by releasing phenolic compounds and producing bioactive peptides, which boost the functional properties of fermented foods (Sumeyye Saritas et al., 2024).

Table No. 2 Comparative Analysis of Controlled and Spontaneous Fermentation Processes

Step	Controlled Fermentation	Spontaneous Fermentation	Reference
Substrate Preparation	Substrate is sterilized or pasteurized to eliminate unwanted microbes, ensuring a selective environment for the desired strains	Substrate is often raw or minimally processed, allowing indigenous microbes to initiate fermentation	(Chrysa Voidarou et al., 2020; R.B. Cuvas – Limon et al., 2020)
Inoculation	Specific microbial strains (e.g., <i>Lactobacillus plantarum</i> , <i>Saccharomyces cerevisiae</i>) are	No inoculum is added; fermentation is initiated by native microflora present in	(Jyoti Prakash Tamang et al., 2020; R.B.

	deliberately introduced to control fermentation	the environment or substrate	Cuvas–Limon et al., 2020)
Fermentation Conditions	Strict control over temperature, pH, oxygen levels, and time ensures reproducibility and safety	Environmental conditions vary, leading to inconsistent fermentation rates and microbial profiles	(Jyoti Prakash Tamang et al.,2020; R.B. Cuvas – Limon et al., 2020)
Microbial Succession	Predictable microbial succession due to defined inocula and controlled settings.	Unpredictable succession due to diversity and competition among indigenous microbes	(Jyoti Prakash Tamang et al.,2020; R.B. Cuvas–Limon et al., 2020)
Monitoring and Control	Progress is monitored using biochemical and molecular tools; deviations can be corrected in real-time	Minimal or no monitoring; fermentation evolves naturally, often without corrective intervention	(Chrysa Voidarou et al., 2020; R.B. Cuvas – Limon et al., 2020)
End Product Characteristics	Result in consistent sensory profiles, quality, and microbiological safety	Product quality varies due to microbial diversity and environmental fluctuations	(Jyoti Prakash Tamang et al.,2020)

Controlled Fermentation utilizes specific starter cultures to offer constant product quality and safety. The process allows predictable fermentation results and is normally followed in industrial food processing (Jyoti P Tamang et al., 2016). Spontaneous fermentation favors the growth of yeasts and bacteria that transform the substrate (sugar-rich) into an alcoholic drink. This fermented product contains nutritional compounds like amino acids, proteins, vitamins, and sugar (A.C. Flores-Gallegos et al.,2019).

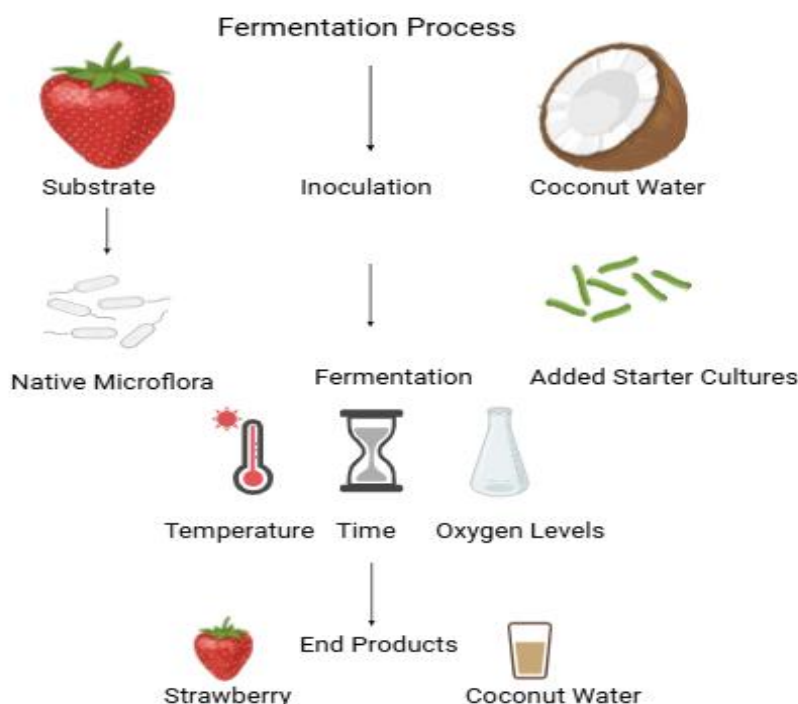


Figure 1: Fermentation in Strawberry and Coconut Water

5. Exploring Microbial Diversity During the Fermentation of Coconut and Strawberry Products

Table No. 3 Comparative Microbial Profile and Identification Techniques in Coconut and Strawberry Fermentation.

Aspect	Coconut Fermentation	Strawberry Fermentation	Type	References
Indigenous Microorganisms	LAB (e.g., <i>L. plantarum</i>) naturally present	Actinobacteria, etc. from natural microbiota	Both fermentations are spontaneous	(Davani Pavalakumar et al. 2024; Raghuram Badmi et al., 2023)
Introduced Microorganisms	<i>L. reuteri</i> added for reuterin production	<i>S. cerevisiae</i> , <i>Lactobacillus spp.</i> for flavor/preservation	Both fermentations are controlled	(Carolina Saori Ishii Mauro et al., 2019; Xinxing Xu et al., 2018)
Spoilage Organisms	Potential contamination if not controlled	Spoilage from <i>Pseudomonas spp.</i> , <i>Botrytis</i> etc.	Both fermentations are spontaneous	Raghuram Badmi et al., 2023)
Identification Methods (Conventional)	Biochemical tests	Similar methods for initial identification	Both are Controlled and Spontaneous fermentation	(Mansi Limbad et al., 2024)
Identification methods (Molecular)	16S rRNA, Illumina MiSeq	High-throughput sequencing	Both fermentations are controlled	(Mansi Limbad et al., 2024; Raghuram Badmi et al., 2023)

This table summarizes the complementary dualities of fermentation: native microbiota-driven spontaneous fermentation and controlled fermentation typified by deliberately introducing specific advantageous microorganisms.

6. Challenges Affecting Quality and Stability in Coconut and Strawberry Ferments

Strawberry-derived fermented products are severely challenged by the high perishability of the fruit, which results in the quick depletion of volatile compounds and nutritional constituents during processing. Thermovinification, although a good method for flavor enhancement and color extraction, has been found to break down essential bioactive compounds like quercetin, ellagic acid, vitamin C, anthocyanins, and other phenolics. Furthermore, alcoholic fermentation is responsible for a loss of 19% of anthocyanins, and storage leads to retention of only 1–2% monomeric anthocyanins. Furthermore, high acetic acid levels also have detrimental effects on the overall aroma of the final product (Bezerra et al., 2024). In coconut fermentations, some issues are high microbial counts in spontaneous fermentation (no added yeast), which raises the risk of contamination. A significant decrease in pH (as low as 2.4) has been reported, which may prevent yeast activity and destabilize fermentation. In addition, high-speed sugar utilization during fermentation can modify alcohol yield and sensory properties, impacting the overall quality of the product (C.O. Ajogun et al., 2020). These issues in combination affect the nutritional, sensory, and microbial stability of strawberry and coconut fermented products. Hence, the use of traditional fermentation control techniques—like defined starter cultures, pH control, and controlled fermentation parameters—is necessary to improve product consistency, safety, and functional value.

7. Conclusion

Controlled fermentation is the most significant factor in the improvement of the nutritional, sensory, and functional attributes of coconut- and strawberry-derived beverages. The disadvantages of spontaneous fermentation—microbial instability, nutrient loss, and variable sensory quality—underline the significance of controlled fermentation methods. Using the application of chosen microbial strains, pH control, and the inclusion of kinetic modelling, stable and reproducible manufacture of beverages is possible. Strawberries and coconuts, high in bioactive compounds, are ideal substrates to produce functional foods. Upon fermentation in well-controlled processes, these fruits yield products containing intact essential vitamins, antioxidants, and phytochemicals, yielding longer shelf life, taste, and therapeutic activity. The intersection of reproducibility, health value, and clean-label acceptance fits into modern food trends and the advancement in fermentation science. Lastly, controlled fermentation is a bridge between traditional practice and contemporary functional food technology for producing safe, health-enhancing, and economically viable coconut and strawberry beverages.

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References

1. Ajogun, C. O., Achinewhu, S. C., Kiin-Kabari, D. B., & Akusu, O. M. (2020). Physicochemical, sensory and microbiological quality of table wine produced from coconut water, honey and zobo. *European Journal of Agriculture and Food Sciences*, 2(5).
2. Albergaria, H., & Arneborg, N. (2016). Dominance of *Saccharomyces cerevisiae* in alcoholic fermentation processes: role of physiological fitness and microbial interactions. *Applied microbiology and biotechnology*, 100, 2035-2046.
3. Badmi, R., Gogoi, A., & Doyle Prestwich, B. (2023). Secondary Metabolites and Their Role in Strawberry Defense. *Plants (Basel, Switzerland)*, 12(18), 3240. <https://doi.org/10.3390/plants12183240>
4. Behera, S. S., Ray, R. C., & Zdolec, N. (2018). *Lactobacillus plantarum* with Functional Properties: An Approach to Increase Safety and Shelf-Life of Fermented Foods. *BioMed research international*, 2018, 9361614. <https://doi.org/10.1155/2018/9361614>
5. Bezerra, M., Ribeiro, M., Cosme, F., & Nunes, F. M. (2024). Overview of the distinctive characteristics of strawberry, raspberry, and blueberry in berries, berry wines, and berry spirits. *Comprehensive reviews in food science and food safety*, 23(3), e13354. <https://doi.org/10.1111/1541-4337.13354>
6. Capece, A., Votta, S., Guaragnella, N., Zambuto, M., Romaniello, R., & Romano, P. (2016). Comparative study of *Saccharomyces cerevisiae* wine strains to identify potential marker genes correlated to desiccation stress tolerance. *FEMS yeast research*, 16(3), fow015.
7. Combina, M., Elía, A., Mercado, L., Catania, C., Ganga, A., & Martinez, C. (2005). Dynamics of indigenous yeast populations during spontaneous fermentation of wines from Mendoza, Argentina. *International journal of food microbiology*, 99(3), 237–243. <https://doi.org/10.1016/j.ijfoodmicro.2004.08.017>
8. Cuarto, P. M., & Magsino, R. F. (2017). Development of young coconut (*Cocos nucifera*) wine. *Asia Pacific Journal of Multidisciplinary Research*, 5(2), 89-93.
9. Cuvás-Limon, R. B., Nobre, C., Cruz, M., Rodriguez-Jasso, R. M., Ruíz, H. A., Loredó-Treviño, A., ... & Belmares, R. (2021). Spontaneously fermented traditional beverages as a source of

- bioactive compounds: an overview. *Critical reviews in food science and nutrition*, 61(18), 2984-3006.
10. De Souza, A. S., Coutinho, J. P., de Souza, L. B. B. C., Barbosa, D. P., da Silva Júnior, A. L. S., & Paixão, M. V. S. (2020). Physical-Chemical Characterization of Fermented Coconut Water (*Cocos nucifera* L.). *Int. J. Adv. Eng. Res. Sci*, 7, 247-255.
 11. Dong, J., Zhang, Y., Tang, X., Jin, W., & Han, Z. (2013). Differences in volatile ester composition between *Fragaria* × *ananassa* and *F. vesca* and implications for strawberry aroma patterns. *Scientia Horticulturae*, 150, 47-53.
 12. Doriya, K., Kumar, D. S., & Thorat, B. N. (2022). A systematic review on fruit-based fermented foods as an approach to improve dietary diversity. *Journal of Food Processing and Preservation*, 46(11), e16994.
 13. Fischer, U., Strasser, M., & Gutzler, K. (2000). Impact of fermentation technology on the phenolic and volatile composition of German red wines. *International Journal of Food Science and Technology*, 35(1), 81-94.
 14. Flores-Gallegos, A. C., Vázquez-Vuelvas, O. F., López-López, L. L., Sainz-Galindo, A., Ascacio-Valdes, J. A., Aguilar, C. N., & Rodriguez-Herrera, R. (2019). Tuba, a fermented and refreshing beverage from coconut palm sap. In *Non-alcoholic beverages* (pp. 163-184). Woodhead Publishing.
 15. Forney, C. F., Kalt, W., & Jordan, M. A. (2000). The composition of strawberry aroma is influenced by cultivar, maturity, and storage. *HortScience*, 35(6), 1022-1026.
 16. Giampieri, F., Alvarez-Suarez, J. M., & Battino, M. (2014). Strawberry and human health: Effects beyond antioxidant activity. *Journal of agricultural and food chemistry*, 62(18), 3867-3876.
 17. Hamilton, P. D., Charles, K. T., Loh, A. M. B., Loïc, N. N. A., Germain, K., & Elie, F. (2024). Physicochemical, nutritional, antioxidant properties and stability monitoring of coconut (*Cocos nucifera* L.) water from two localities in Cameroon. *Heliyon*, 10(23).
 18. Hornedo-Ortega, R., Álvarez-Fernández, M. A., Cerezo, A. B., Garcia-Garcia, I., Troncoso, A. M., & Garcia-Parrilla, M. C. (2017). Influence of Fermentation Process on the Anthocyanin Composition of Wine and Vinegar Elaborated from Strawberry. *Journal of food science*, 82(2), 364–372. <https://doi.org/10.1111/1750-3841.13624>
 19. Jagadeesh, U., Narayanaswamy, B., & Suvarna, V. C. (2019). *Microbial processing of coconut water for the development of coconut wine* (Doctoral dissertation, University of Agricultural Sciences, GKVK).
 20. Joshi, V. K., & Kumar, V. (2011). Importance, nutritive value, role, present status, and future strategies in fruit wines in India. *Bio-Processing of Foods*. (Eds. PS Panesar et al.), 39-62.
 21. Kochadai, N., Mahendran, R., Bhosale, Y. K., Vincent, H., & Nair, S. V. R. (2021). Development of low alcoholic wine using tender coconut and tender palmyra as a novel source and its quality evaluation.
 22. Lan, W., Zhang, M., Xie, X., Li, R., Cheng, W., Ma, T., & Zhou, Y. (2024). Effects of Cultivar Factors on Fermentation Characteristics and Volatile Organic Components of Strawberry Wine. *Foods (Basel, Switzerland)*, 13(18), 2874. <https://doi.org/10.3390/foods13182874>
 23. Limbad, M., Gutierrez Maddox, N., Hamid, N., Kantono, K., & Higgins, C. (2024). Identification of the Microbiota in Coconut Water, Kefir, Coconut Water Kefir and Coconut Water Kefir-Fermented Sourdough Using Culture-Dependent Techniques and Illumina-MiSeq Sequencing. *Microorganisms*, 12(5), 919. <https://doi.org/10.3390/microorganisms12050919>
 24. Liu, S., Laaksonen, O., Li, P., Gu, Q., & Yang, B. (2022). Use of non-Saccharomyces yeasts in berry wine production: Inspiration from their applications in winemaking. *Journal of Agricultural and Food Chemistry*, 70(3), 736-750.
 25. Lv, Z., Liu, H., Yang, W., Zhang, Q., Chen, D., Jiao, Z., & Liu, J. (2024). Comprehensive Analysis of Physicochemical Properties and Volatile Compounds in Different Strawberry Wines under Various Pre-Treatments. *Molecules (Basel, Switzerland)*, 29(9), 2045. <https://doi.org/10.3390/molecules29092045>

26. Maicas, S. (2020). The role of yeasts in fermentation processes. *Microorganisms*, 8(8), 1142.
27. Mani, A. (2018). Food preservation by fermentation and fermented food products. *Int. J. Acad. Res. Dev*, 1, 51-57.
28. Mauro, C. S. I., & Garcia, S. (2019). Coconut milk beverage fermented by *Lactobacillus reuteri*: optimization process and stability during refrigerated storage. *Journal of food science and technology*, 56(2), 854–864. <https://doi.org/10.1007/s13197-018-3545-8>
29. Mauro, C. S. I., Fernandes, M. T. C., Farinazzo, F. S., & Garcia, S. (2022). Characterization of a fermented coconut milk product with and without strawberry pulp. *Journal of food science and technology*, 59(7), 2804–2812. <https://doi.org/10.1007/s13197-021-05303-1>
30. Mohammed, S. S. D., Yohanna, B., Wartu, J. R., Abubakar, N. L., & Bello, S. (2018). Wine produced from fermentation of honey slurry and ddates palm fruit juice blend using *Saccharomyces cerevisiae* isolated from palm wine. *Int. J. Biol*, 10(3), 52.
31. Morata, A., Arroyo, T., Bañuelos, M. A., Blanco, P., Briones, A., Cantoral, J. M., Castrillo, D., Cordero-Bueso, G., Del Fresno, J. M., Escott, C., Escribano-Viana, R., Fernández-González, M., Ferrer, S., García, M., González, C., Gutiérrez, A. R., Loira, I., Malfeito-Ferreira, M., Martínez, A., Pardo, I., ... Capozzi, V. (2023). Lan, W., Zhang, M., Xie, X., Li, R., Cheng, W., Ma, T., & Zhou, Y. (2024). Effects of cultivar factors on fermentation characteristics and volatile organic components of strawberry wine. *Foods*, 13(18), 2874.
32. Newerli-Guz, J., Śmiechowska, M., Drzewiecka, A., & Tylingo, R. (2023). Bioactive ingredients with health-promoting properties of strawberry fruit (*Fragaria x ananassa* Duchesne). *Molecules*, 28(6), 2711.
33. Pavalakumar, D., Undugoda, L. J. S., Gunathunga, C. J., Manage, P. M., Nugara, R. N., Kannangara, S., Lankasena, B. N. S., & Patabendige, C. N. K. (2024). Evaluating the Probiotic Profile, Antioxidant Properties, and Safety of Indigenous *Lactobacillus* spp. Inhabiting Fermented Green Tender Coconut Water. *Probiotics and antimicrobial proteins*, 10.1007/s12602-024-10352-x. Advance online publication. <https://doi.org/10.1007/s12602-024-10352-x>
34. Perez, A. G., Rios, J. J., Sanz, C., & Olias, J. M. (1992). Aroma components and free amino acids in the strawberry variety Chandler during ripening. *Journal of Agricultural and Food Chemistry*, 40(11), 2232-2235.
35. Rabitti, N. S., Cattaneo, C., Appiani, M., Proserpio, C., & Laureati, M. (2022). Describing the sensory complexity of Italian wines: Application of the Rate-All-That-Apply (RATA) method. *Foods*, 11(16), 2417.
36. Raymond Eder, M. L., & Rosa, A. L. (2021). Non-conventional grape varieties and yeast starters for first and second fermentation in sparkling wine production using the traditional method. *Fermentation*, 7(4), 321.
37. Rocha, B. P., de Brito Lopes, P. L., da Silva, M. O. M., Gomes, A. C. G., Buriti, F. C. A., Florêncio, I. M., & Florentino, E. R. (2024). Utilization of ripe coconut water in the development of probiotic gelatin. *PeerJ*, 12, e17502.
38. Romano, P., Siesto, G., Capece, A., Pietrafesa, R., Lanciotti, R., Patrignani, F., Granchi, L., Galli, V., Bevilacqua, A., Campaniello, D., Spano, G., Caridi, A., Poiana, M., Foschino, R., Vigentini, I., Blaiotta, G., Corich, V., Giacomini, A., Cardinali, G., Corte, L., ... Tufariello, M. (2022). Validation of a Standard Protocol to Assess the Fermentative and Chemical Properties of *Saccharomyces cerevisiae* Wine Strains. *Frontiers in microbiology*, 13, 830277. <https://doi.org/10.3389/fmicb.2022.830277>
39. Ross, R. P., Morgan, S., & Hill, C. (2002). Preservation and fermentation: past, present and future. *International journal of food microbiology*, 79(1-2), 3–16. [https://doi.org/10.1016/s0168-1605\(02\)00174-5](https://doi.org/10.1016/s0168-1605(02)00174-5)
40. Sadik, H., Ouazzani, C., Moustaghfir, A., El Ghammarti, S., Er-Ramly, A., Essebbahi, I., ... & Balouch, L. (2023). Comparison of the nutritional proprieties of commercial strawberries, red and black raspberries consumed in Morocco. *Applied Food Research*, 3(2), 100362.

41. Sáenz-Navajas, M. P., Sánchez, C., Gonzalez-Hernandez, M., Bueno, M., Peña, C., Fernández-Zurbano, P., Ballester, J., Parga-Dans, E., & González, P. A. (2023). Natural versus conventional production of Spanish white wines: an exploratory study. *Journal of the science of food and agriculture*, 103(7), 3540–3549. <https://doi.org/10.1002/jsfa.12479>
42. Şanlıer, N., Gökçen, B. B., & Sezgin, A. C. (2019). Health benefits of fermented foods. *Critical reviews in food science and nutrition*, 59(3), 506–527. <https://doi.org/10.1080/10408398.2017.1383355>
43. Sarıtaş, S., Portocarrero, A. C. M., Miranda López, J. M., Lombardo, M., Koch, W., Raposo, A., El-Seedi, H. R., de Brito Alves, J. L., Esatbeyoglu, T., Karav, S., & Witkowska, A. M. (2024). The Impact of Fermentation on the Antioxidant Activity of Food Products. *Molecules (Basel, Switzerland)*, 29(16), 3941. <https://doi.org/10.3390/molecules29163941>
44. Sharma, S., Joshi, V. K., & Abrol, G. (2009). An overview on strawberry [*Fragaria×ananassa* (Weston) Duchesne ex Rozier] wine production technology, composition, maturation, and quality evaluation.
45. Sreelekshmi, M. M. R., Sayoojya, K. P., Souparnika, A. P., Sowparnika, K., Pournami, T. S., Sabu, K. R., ... & Chandran, R. P. (2018). Production of coconut sprout wine using *Saccharomyces cerevisiae* and its physico-chemical analysis. *MOJ Food Process Technol*, 6(5), 445-449.
46. Tamang, J. P., Cotter, P. D., Endo, A., Han, N. S., Kort, R., Liu, S. Q., ... & Hutkins, R. (2020). Fermented foods in a global age: East meets West. *Comprehensive Reviews in Food Science and Food Safety*, 19(1), 184-217.
47. Tamang, J. P., Shin, D. H., Jung, S. J., & Chae, S. W. (2016). Functional Properties of Microorganisms in Fermented Foods. *Frontiers in microbiology*, 7, 578. <https://doi.org/10.3389/fmicb.2016.00578>
48. Tofalo, R., Perpetuini, G., Rossetti, A. P., Gaggiotti, S., Piva, A., Olivastri, L., ... & Arfelli, G. (2022). Impact of *Saccharomyces cerevisiae* and non-*Saccharomyces* yeasts on improving traditional sparkling wines production. *Food Microbiology*, 108, 104097.
49. Tulipani, S., Mezzetti, B., & Battino, M. (2009). Impact of strawberries on human health: insight into marginally discussed bioactive compounds for the Mediterranean diet. *Public health nutrition*, 12(9A), 1656–1662. <https://doi.org/10.1017/S1368980009990516>
50. Udomsaksakul, N., Kodama, K., Tanasupawat, S., & Savarajara, A. (2018). Indigenous *Saccharomyces cerevisiae* strains from coconut inflorescence sap: characterization and use in coconut wine fermentation. *CMU J Nat Sci*, 17(3), 219-230.
51. Vilela, A. (2017). Biological demalization and deacetication of musts and wines: Can wine yeasts make the wine taste better?. *Fermentation*, 3(4), 51.
52. Voidarou, C., Antoniadou, M., Rozos, G., Tzora, A., Skoufos, I., Varzakas, T., ... & Bezirtzoglou, E. (2020). Fermentative foods: Microbiology, biochemistry, potential human health benefits and public health issues. *Foods*, 10(1), 69.
53. Xu, X., Luo, D., Bao, Y., Liao, X., & Wu, J. (2018). Characterization of Diversity and Probiotic Efficiency of the Autochthonous Lactic Acid Bacteria in the Fermentation of Selected Raw Fruit and Vegetable Juices. *Frontiers in microbiology*, 9, 2539. <https://doi.org/10.3389/fmicb.2018.02539>
54. Yang, W., Liu, S., Marsol-Vall, A., Tähti, R., Laaksonen, O., Karhu, S., ... & Ma, X. (2021). Chemical composition, sensory profile and antioxidant capacity of low-alcohol strawberry beverages fermented with *Saccharomyces cerevisiae* and *Torulaspora delbrueckii*. *Lwt*, 149, 111910.
55. Yong, J. W. H., Ge, L., Ng, Y. F., & Tan, S. N. (2009). The Chemical Composition and Biological Properties of Coconut (*Cocos nucifera* L.) Water. *Molecules*, 14(12),
56. Yuan, Y. H., Mu, D. D., Guo, L., Wu, X. F., Chen, X. S., & Li, X. J. (2024). From flavor to function: A review of fermented fruit drinks, their microbial profiles and health benefits. *Food Research International*, 115095.

57. Zabetakis, I., & Holden, M. A. (1997). Strawberry flavour: analysis and biosynthesis. *Journal of the Science of Food and Agriculture*, 74(4), 421-434.
58. Zhang, G., Chen, W., Chen, W., & Chen, H. (2018). Improving the quality of matured coconut (*Cocos nucifera* Linn.) water by low alcoholic fermentation with *Saccharomyces cerevisiae*: antioxidant and volatile profiles. *Journal of food science and technology*, 55(3), 964–976. <https://doi.org/10.1007/s13197-017-3004-y>
59. Zhang, G., Li, X., Chen, W., Chen, P., Jin, X., Chen, W., & Chen, H. (2018). Organic acid content, antioxidant capacity, and fermentation kinetics of matured coconut (*Cocos nucifera*) water fermented by *Saccharomyces cerevisiae* D254. *International Journal of Food Engineering*, 14(3), 20170331
60. Zhang, Y., Kan, J., Liu, X., Song, F., Zhu, K., Li, N., & Zhang, Y. (2024). Chemical components, nutritional value, volatile organic compounds and biological activities in vitro of coconut (*Cocos nucifera* L.) water with different maturities. *Foods*, 13(6), 863.
61. Zhao, Z., Wu, X., Chen, H., Liu, Y., Xiao, Y., Chen, H., Tang, Z., Li, Q., & Yao, H. (2021). Evaluation of a strawberry fermented beverage with potential health benefits. *PeerJ*, 9, e11974. <https://doi.org/10.7717/peerj.11974>