



## TOPICAL GELS FOR DRUG DELIVERY: RECENT ADVANCEMENTS IN FORMULATION AND FUTURE DIRECTIONS IN CLINICAL PRACTICE

Sadia Ahmed Zuberi<sup>1\*</sup>, Fawaz Jaffar<sup>1</sup>, Nousheen Alam<sup>1</sup>, Samina Sheikh<sup>2</sup>, Nimra Aamir<sup>1</sup>,  
Hasnan Ali<sup>3</sup>, Rahat Fatima Naqvi<sup>4</sup>

<sup>1\*</sup>Faculty of Pharmaceutical Sciences, SBB Dewan University, Karachi, Pakistan

<sup>2</sup>Faculty of Pharmacy and Pharmaceutical Sciences, Ziauddin University, Karachi, Pakistan

<sup>3</sup>Product Lead, Manufacturing Sciences and Technology, GlaxoSmithKline, Karachi, Pakistan

<sup>4</sup>Faculty of Pharmacy, IBADAT International University, Islamabad

**\*Corresponding author:** Sadia Ahmed Zuberi  
email: rphsadiazuberi@gmail.com

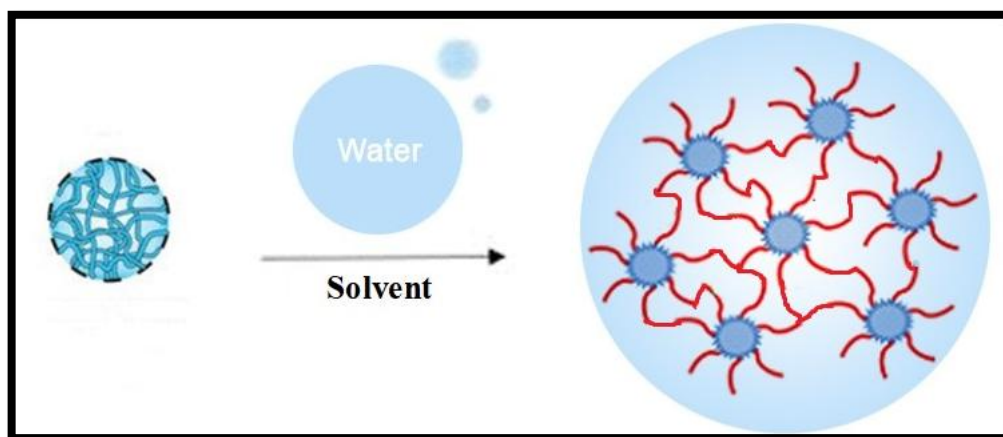
### Abstract

The main intention behind this review is to provide updated information related to the recent developments in terms of novel topical preparations including the basic concepts of dermal drug delivery systems. Topical drug delivery systems especially gels have several advantages such as it provides prolonged application to the site of action ultimately improving the therapeutic effect. A gel is a semisolid combination of a three-dimensional matrix of cross-linked materials. The three dimensional matrix of gel is mainly composed of a gelling agent and water. This review highlights the fundamentals of gel formulation and the factors affecting, along with the classification, advantages, limitations, and applications of different types of pharmaceutical gels such as niosomal/proniosomal gel, emulgels, bigels, xerogels and aerogels, organogels and hydrogels.

**Keywords:** Classification, Gel applications, gel formulation, limitations, pharmaceutical gels.

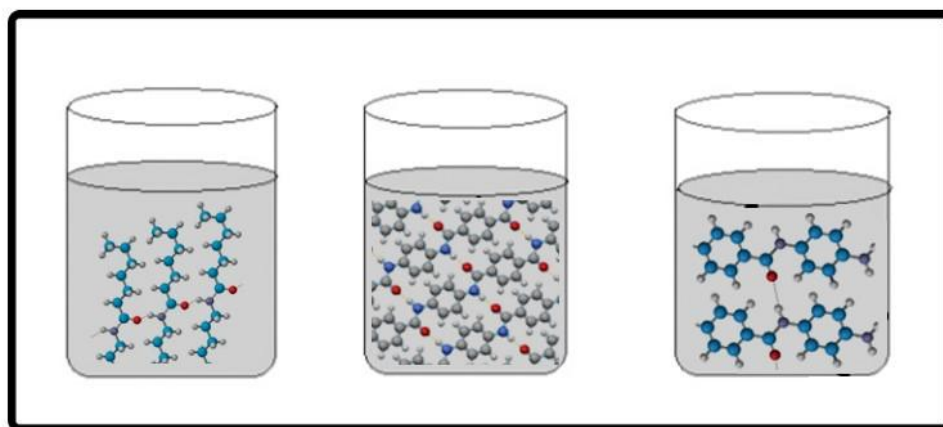
### 1. INTRODUCTION

A gel is a semisolid combination of a three-dimensional matrix of cross-linked materials. Several substances create a matrix in a liquid dispersion medium and form a gel (Fig. 1.1.). Organic and inorganic macromolecules act as formational units in preparing gel (Larson, 1999). According to USP (2021), "A gel is solid or semisolid. Gels can be classified into two groups, chemical and physical gels. Chemical gels are usually covalently cross-linked gels, while physical gels consist of small molecules or molecular chains that are physically cross-linked into networks, or solutions, or colloidal dispersions that are stiffened by a gelling agent." A gel is a two-phase system in which the inorganic particles are present in an insoluble state, but these particles are uniformly distributed throughout the external phase. However, the large organic particles are freely available in the external phase (Fig. 1.1.). These phases are then joined together in stretchable chains (Goyal et al., 2011).



**Fig. 1.1.** Cross-linking of polymer in a solvent to form a gel matrix.

Gelling agent, also known as a gelator, is a natural, synthetic or semi-synthetic polymer or low molecular weight small molecule which is added to an organic, inorganic, or aqueous solvent or in a solvent system to prepare a gel (Fig. 1.2) (Jain, 2006; Murdan, 2005; Vintiloiu and Leroux, 2008; Allan, 2012).



**Fig. 1.2.** Swelling of a polymer to form a gel.

## 1.2. TECHNICAL FEATURES OF GELS

Gels have been the topic of interest for the past few decades because of their potential for application in specific areas such as biomedical, cosmeceutical, energy-saving, etc. One of the recent features focused on is the adhesiveness of the gel on different surfaces (Sun et al., 2021). Some of the technical features that are important for gels are discussed below:

- i. The gelling agent should be inert and should not produce any harmful effect on the formulation.
- ii. Anti-microbial agents should be present in the formulation to stabilize it from microbial contamination.
- iii. It should be safe and provide patient compliance.
- iv. All rheological properties of the gel should be maintained.
- v. The gel formulation should be economical.
- vi. Gel intended to be used in eyes should be sterile and stored at the recommended storage condition to maintain stability.
- vii. It should be neither greasy nor sticky and non-staining in nature.
- viii. No physical change should occur in the formulation upon storage.
- ix. It should have the capability of a high level of drug encapsulation.
- x. The gel must be biodegradable without forming toxins.

### 1.3. ADVANTAGES OF GELS OVER OTHER DOSAGE FORMS

Recent advances in gel technology have proven crucial for their application, especially in topical drug delivery. Some of the advantages of gels that make them superior to other dosage forms are enlisted here (Jain, 2006; Florence and Attwood, 2007; Labarre et al., 2010; Rehman et al., 2014):

- i. They produce a local effect; therefore, there are fewer chances of side effects.
- ii. They provide a soothing effect on the skin.
- iii. They are less greasy as compared to other dosage forms and easily washable.
- iv. They are non-invasive and have a high retention time compared to other dermatological preparations, thus providing patient compliance.
- v. They provide targeted and controlled drug delivery to the diseased tissue with the advantage of tissue repair by releasing the drug through chemical stimulation.
- vi. They can incorporate hydrophilic as well as lipophilic drugs.

### 1.4. GEL LIMITATIONS / DISADVANTAGES

Apart from being advantageous, the gels also have some limitations, which are discussed as follows (Nabi et al., 2016):

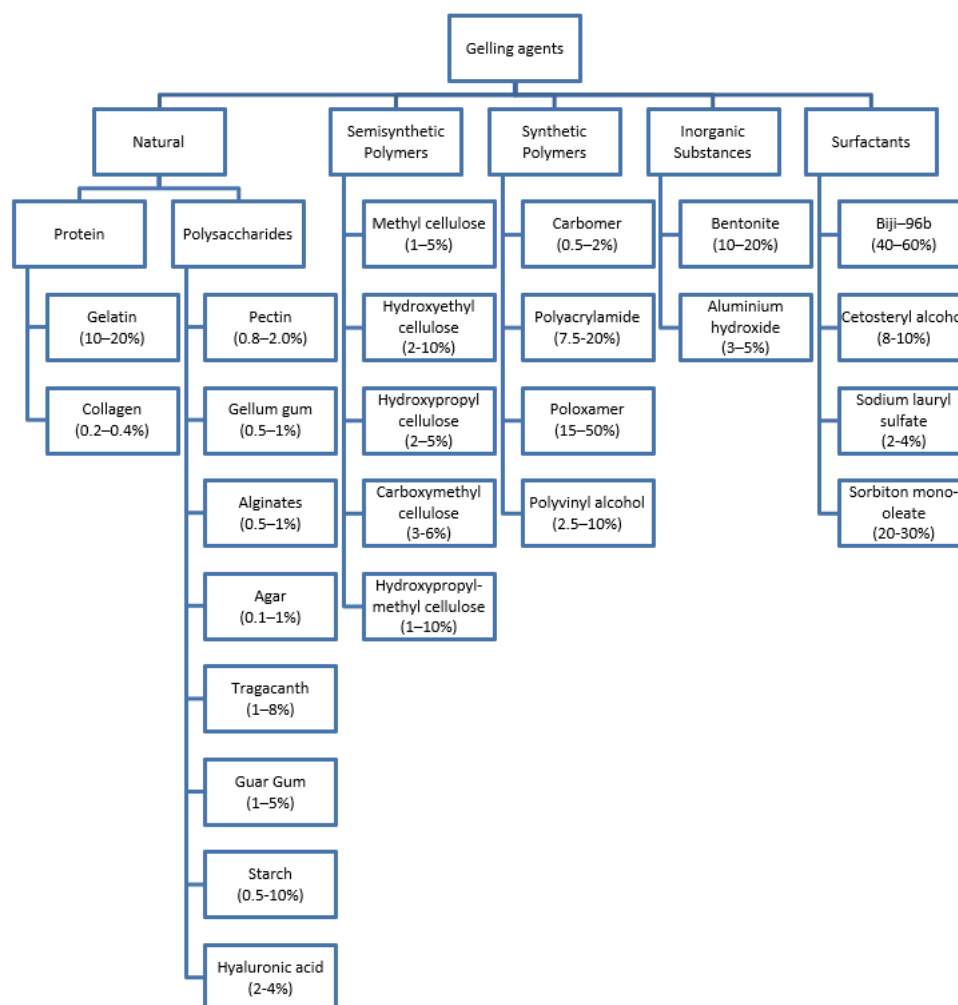
- i. If a drug with a large particle size is incorporated into the gel, it becomes difficult to get absorbed through the skin.
- ii. Topical gels can cause allergic reactions.
- iii. A high quantity of water may result in microbial or fungal growth.
- iv. Syneresis is another issue that may occur on storage.
- v. The release of the drug may be affected in gels with a covalent bond.
- vi. Temperature, humidity, and other environmental factors may alter the rheological properties of the gels.
- vii. Degradation of a drug may result from polymer interaction with the drug in some cases.

### 1.5. BASIC COMPONENTS OF A GEL FORMULATION

The main advantage of formulating a gel is the ease of preparation. The fundamental components include a polymer (gelling agent) and a solvent. Apart from these two, a gel may contain stabilizing agents, dispersing agents, buffer, preservatives, penetration enhancers, anti-oxidants, and flavoring agents/sweeteners. Preservatives are essential as they prevent the gel from microbial growth, especially in the case of hydrogel, due to the presence of a high amount of water (Nabi et al., 2016). Anti-oxidants are added in preparations that are sensitive to oxidative degradation, and their selection depends on the type of solvent used. Water-soluble anti-oxidants (e.g., sodium metabisulfite, sodium formaldehyde sulfoxylate, etc.) are preferred as a majority of the gels have an aqueous solvent (Rathod and Mehta, 2015).

#### 1.5.1. Gelling Agents

The selection of a gelling agent depends on the gel type (e.g., hydrogel, organogel, etc.) and its affinity with the solvent. Fig. 1.3 enlists some of the commonly used gelling agents with their classification and percentages at which they act as a gelling agent (Attwood., 2002; Rathod and Mehta, 2015).



**Fig. 1.3.** The classification of gelling agents with origin and percentages.

### 1.5.2. Buffers

pH plays a vital role in the penetration of the drug through the skin. Buffers may be added, especially in topical drug delivery-based gels. The buffer selection is critical as it has some hydro-alcoholic-based solvents limitations. Examples of buffers include phosphate buffer, citrate buffer, etc. (Bhatia et al., 2000; Rathod and Mehta, 2015).

### 1.5.3. Penetration Enhancers

The penetration enhancer should be inert chemically and pharmacologically but must be acceptable pharmaceutically. They should be free from allergic reactions and non-irritant, colorless, odorless, and tasteless, having a known duration of action with a quick response. More importantly, it must be accepted cosmeceutically (Ojha et al., 2017).

### 1.5.4. Gel Preparation Medium

A solvent is often purified water but co-solvents (e.g., alcohol, glycerol, PEG, etc.) may be added to enhance drug solubility or increase the drug skin penetration (Rathod et al., 2015). In nature, a solvent can be hydrophilic or hydrophobic (lipophilic or organic). The selection of the vehicle-like gelling agent depends on the desired application of the gel. The solvent selected must be efficient, biocompatible, and not evaporate quickly (Nabi et al., 2016).

## 1.6. FACTORS AFFECTING GEL FORMULATIONS

Formulating a gel is easy as compared to other dosage forms. However, certain factors affect the gel formulations, which include (Nabi et al., 2016):

- i. Gelling agent concentration and its molecular weight.
- ii. Solubility and affinity of the polymer towards the solvent used.
- iii. pH.
- iv. Solvent nature.
- v. Ion concentration of the solution.
- vi. Temperature, humidity, and other environmental factors.

## 1.7. STRUCTURE AND CLASSIFICATION OF GELS

In recent drug developments, gels have become the most beneficial vehicle for localized, topical drug delivery. Advanced studies for the dermal application of the drug have reported some additional types of gels (i.e., niosomal gel, emulgels, bigels, xerogels), including hydrogels and oleogels (Rehman et al., 2014).

Conventional methods of gel formation are cost-effective and convenient to make. However, the advancement in the field of the gel makes it challenging to characterize the gel, especially the hydrogels having a variety of cross-linked networks (Peppas and Barr-howell, 2019). Khandan et al. (2017) characterized the gels as liquids having 3D cross-linked structure which behaves as solids. The interlinking of the fragments of the gelling agent strengthens the gel. Particle nature and the type of force involved in connecting these linkages relate to the gel's structure and properties. The attractive force involved in the pairing of gelling agents may vary from strong covalent or ionic bonds to weak hydrogen bonds and Vander Waal forces. A slight rise in temperature occurs due to liquefaction in gels due to weak interactions (Carter, 2000). A brief classification of gels is elucidated in Table 1.1.

**Table 1.1.** Classification of gels

Physical Properties	<ul style="list-style-type: none"> <li>• Smart</li> <li>• Conventional</li> </ul>
Ionic Charge	<ul style="list-style-type: none"> <li>• Non-ionic</li> <li>• Cationic</li> <li>• Anionic</li> <li>• Ampholytic</li> </ul>
Cross-linking	<ul style="list-style-type: none"> <li>• Physical</li> <li>• Chemical</li> </ul>
Biodegradability	<ul style="list-style-type: none"> <li>• Biodegradable</li> <li>• Non-biodegradable</li> </ul>
Size of Pores	<ul style="list-style-type: none"> <li>• Non-porous</li> <li>• Microporous</li> <li>• Nanoporous</li> <li>• Macroporous</li> </ul>
Preparation Method	<ul style="list-style-type: none"> <li>• Homopolymer</li> <li>• Copolymer: Triblock and Multiblock</li> <li>• Interpenetrating</li> </ul>
Source	<ul style="list-style-type: none"> <li>• Natural</li> <li>• Hybrid</li> <li>• Synthetic</li> </ul>

### 1.7.1. Niosomal or Proniosomal Gel

Niosomes are chemically stable, derived from liposomes having nonionic surfactants, capable of carrying both types of drugs, i.e., hydrophilic or lipophilic. They are more stable than liposomes and phospholipid carriers whereas proniosomes are more stable than niosomes (Masotti et al., 2010; Marianecci et al., 2011; Verma et al., 2020). Niosomes are easy to prepare and can also be formed through hydration from proniosomes which are niosomal hybrids having compact crystalline structures (El-Laithy et al., 2011). Apart from various advantages, niosomes are expensive, sensitive to moisture (hydrolysis), and quickly aggregate and sediment (Rehman et al., 2014). The targeted drug delivery of lipid-soluble vitamins and the non-invasive vaccines have been observed through niosome gels prepared by the oleogel technique (Gupta et al., 2005; Gonnet et al., 2010). In contrast, the hydrogel-based niosomes and proniosomes have been effective for the transdermal delivery of the drug (Marianecci et al., 2011; Shehata et al., 2021). Niosomal gels have several advantages over liposomes, such as improved skin permeation with controlled release of the drug, improved stability chemically as well as physically, and are cost-effective in production (Biswal et al., 2008; Mahale et al., 2012; Verma et al., 2020). To release the drug and permeate it from the skin, proniosome requires hydration; that is why dry niosomes are another name given to proniosome (El-Laithy et al., 2011). Comparatively, niosomal hydrogels are more physically stable than proniosomes and have more drug penetration in the transdermal drug delivery system providing a unique vesicle for delivery (Ibrahim et al., 2005; Rehman et al., 2014; Shah et al., 2021).

### 1.7.2. Emulgels

Despite recent developments in novel drug delivery systems, the delivery of lipophilic drugs has always been an area of concern. The answer to the question has been the emulgel, which converts water-in-oil emulsion to an emulgel by adding a gelling agent in the water phase. Both the emulgels, i.e., water-in-oil and oil-in-water, have been used to deliver the lipophilic drugs (Dhawas et al., 2020; Harshitha et al., 2020; Koshani et al., 2021; Salabat and Parsi., 2021). Emulgels also have various other advantages which are favorable dermatologically, such as:

- i. Biofriendly
- ii. Less greasy
- iii. Greater spreadability
- iv. Easy to remove
- v. Improved shelf-life
- vi. High patient acceptability due to the pleasant appearance
- vii. Provides controlled release of drug
- viii. Easy to prepare and are cost-effective

The only disadvantage of emulgel is skin irritation due to the presence of the surface-active agent(s). However, air bubbles can be formed in preparing an emulgel which are settled on standing (Rehman et al., 2014; Pawbake et al., 2020; Charyulu et al., 2021).

The novel topical drug delivery system has reached a new level as the researchers have moved to the advanced form of emulgels, i.e., microemulsion-based gels (MBGs) and nanoemulsion-based gels (NBGs) for the drug delivery. The advantages of MBGs and NBGs, apart from the one listed above, include their particle size, improved thermodynamic stability, targeted drug delivery, and avoidance of the first-pass effect (Chellapa et al., 2015; Choudhry et al., 2017).

### 1.7.3. Bigels

Bigels, as the name indicates, are a combination of hydrogel and oleogel (organogel), but they are free from any surfactant or stabilizer. They are prepared by mixing hydrogel and organogel at a high shear rate (rpm) (Di Michele et al., 2014; Lupi et al., 2016; Shakeel et al., 2018; Shakeel et al., 2019; Martins et al., 2020; Sreekumar et al., 2020). This homogenous system of bigels possesses properties of both gels and provides synergistic effects leading to increased drug penetration. The combined features of hydrogel and organogel make it unique with certain advantages such as they

are easy to prepare, provide a synergistic effect, are free from surfactant; therefore, no skin irritation is reported due to surfactant, and more importantly, they can deliver both types of drug, i.e., lipophilic and hydrophilic. However, sometimes it is not easy to maintain phase separation since the preparation is surfactant-free (Rehman et al., 2014). Moreover, the synergistic effect of the bigels increases its importance in the cosmetic and pharmaceutical industries (Rehman and Zulfakar, 2014; Rehman et al., 2014; Singh et al., 2014; Lupi et al., 2016). In the past few decades, bigels have been studied extensively for the topical delivery of drugs such as metronidazole, ciprofloxacin, ketoprofen, flurbiprofen, tenofovir, maraviroc, etc. (Andonova et al., 2017; Charyulu et al., 2018; Shakeel et al., 2018; Ilomuanya et al., 2020).

#### **1.7.4. Xerogels and Aerogels**

Aerogels and xerogels, commonly known as inorganic gels (because they are made up of silica), have also been studied extensively for the topical application of various drugs, especially for subcutaneous delivery (Uros et al., 2007; Guenther et al., 2008). Apart from the drying procedures, both the aerogels and xerogels are prepared by the sol-gel method. Xerogels are prepared by drying the wet silica gel at standard pressure. In contrast, the aerogels are prepared by avoiding the shrinkage at the supercritical level and preserving the porous structure, which ultimately forms aerogels. Compared with xerogels, the former is more effective because it has more drug solubility leading to increased bioavailability.

Additionally, the release kinetics can be controlled by modifying the aerogels structure using different functional groups. The main difference lies between the inner surface area, i.e., 400–1000 m<sup>2</sup>/g for aerogel and 300–600 m<sup>2</sup>/g for xerogel, and pore size, i.e., the aerogels have larger pores as compared to xerogels (Rehman et al., 2014). Aerogels have various advantages over xerogels. The drug loading at the supercritical stage provides uniform drug distribution at the molecular level, making it easy to incorporate the amorphous form of the drug. This system can also improve the bioavailability of BCS II drugs as the hydrophilic aerogel system collapses in the hydrophilic medium because of internal surface tension within the pores. Both the gels are thermally stable, having low thermal conductivity. The silica aerogels have a restriction that they are not biodegradable. Several studies have used polysaccharides to prepare biodegradable aerogels (Ollio and Ollio, 2009; Alnaief et al., 2011; Cheng et al., 2012; Stergar and Maver, 2016; Ganesan et al., 2018). However, the aerogel may be organic, inorganic, or hybrid; they have proven biocompatible ((Stergar and Maver, 2016).

#### **1.7.5. Organogels**

Organogels, one of the most common and well-known gel forms, contain lipophilic or non-polar liquid as the dispersion medium. Organogels are a network of a three-dimensional thermo-reversible gel consisting of an organogelator and a non-polar solvent. This three-dimensional network is formed by the physical or chemical interaction of gelators when used in less than 15% concentration, preventing the flow of external apolar solvent (Vintiloiu et al., 2008; Sahoo et al., 2011). The fundamental potential reason behind the study of organogels as the drug delivery system is their thermo-reversible property. These gels are thermodynamically stable, biodegradable, biocompatible, and possess high transdermal permeability, but they are greasy and difficult to wash (Murdan., 2005; Rehman et al., 2014). The applications of organogels are broader as they are used in the chemical, pharmaceutical, cosmeceutical, and food industries. Biotechnology is another field in which organogels are used predominantly as a vaccine delivery platform (Esposito et al., 2013, 2016, 2018). Edible oleogels are another interesting application of organogels to the health-promoting food industry, used to prepare different dairy and meat products (Co and Marangoni., 2018; Martin et al., 2020; Puscas et al., 2020).

### 1.7.6. Hydrogels

In the past few decades, synthetic functional polymers have been studied extensively for their biomedical application, especially as hydrogel, to overcome the disadvantages of conventional drug delivery and improve the efficacy of novel drug delivery systems (Kashyap et al., 2005; Vashist et al., 2014). Like organogels, hydrogels are also three-dimensional polymeric structures, but they contain a hydrophilic gelling agent (either synthetic or natural) that can hold a large amount of water (Rehman et al., 2014; Balakrishnan and Jayakrishnan, 2015). The hydrogels are mainly divided into physical and chemical hydrogels depending on the cross-linking. Physical hydrogels are reversible gels since weak bonds form their cross-linking. The chemical hydrogels, also known as permanent gels, are formed by covalent cross-linking (Campoccia et al., 1998; Shapiro, 2011; Allan, 2012; Peppas and Mikos, 2019). They are easy to prepare, cost-effective, biodegradable, adaptable to different compounds, and, more importantly, it serves as a base for other advanced forms of gels such as liposomal gel, emulgels, microgels, nanogels, nanocomposite hydrogels, etc. (Rehman et al., 2014; Khan et al., 2016; Esmaeely-Neisiany et al., 2020). However, it has some limitations towards transdermal drug delivery because of its hydrophilic nature. Microbial contamination is another disadvantage, especially in polysaccharide hydrogels (Rehman et al., 2014). Based on the size of pores, hydrogels can be characterized as non-porous (10–100  $\mu\text{m}$ ), macroporous (0.1–1  $\mu\text{m}$ ), microporous (100–1000  $\mu\text{m}$ ) and nanoporous (<100 nm) (Amin et al., 2009; Khan et al., 2016). Hydrogels are also classified based on cross-linking, i.e., physically cross-linked and chemically cross-linked hydrogels.

Bioadhesive hydrogel is another achievement in the field of hydrogel for novel drug delivery systems. It is formed when one of the phases is biological, resulting in adherence to the two phases and protecting the separation, resulting in increased bioavailability. Kashyap et al. (2005) discussed different routes of administration of bioadhesive hydrogels which are as follows:

- i. Buccal
- ii. Ocular
- iii. Vaginal and cervical
- iv. Peroral

Hydrogels have many applications, including vaccines, stimuli-responsive nanogels, tissue engineering, molecular imprinting, wound dressing, immunoisolation, and novel drug delivery (Kashyap et al., 2005; Ferreira et al., 2013; Hajebi et al., 2019). Nanotechnology has gained importance in the novel drug delivery system in the past few decades, and nanogels have been the focus, especially in transdermal drug delivery (Karg et al., 2019). The uptake of macrophages by nano hydrogels has been demonstrated by Gao et al. (2008), which brings the focus of researchers toward this novel nanohydrogel drug delivery system (Vashist et al., 2014). Nanogels, like hydrogels, consists of a three-dimensional network having physical or chemical cross-linking of the polymer (Soni and Yadav, 2016; Soni et al., 2016; Cuggino et al., 2019; Pinelli et al., 2020; Shah et al., 2020). A summary for all gel types with their definition, advantages, and limitations is given in Table 1.2.

## 1.8. CHARACTERISTICS OF GELS

### 1.8.1. Syneresis

Syneresis can be defined as the oozing of the fluid medium upon standing due to contractions. This phenomenon increases with the decrease in the concentration of the gelling agent and the other way round. It affects the stability of the gel and depicts that the gel became thermodynamically unstable. The contractions also cause the loosening of elastic stress during gel development. As soon as the stresses are mitigated, shortening of the space takes place within the solvent, which exudates the fluid (Carter, 2000; Zatz and Kushla, 2005).

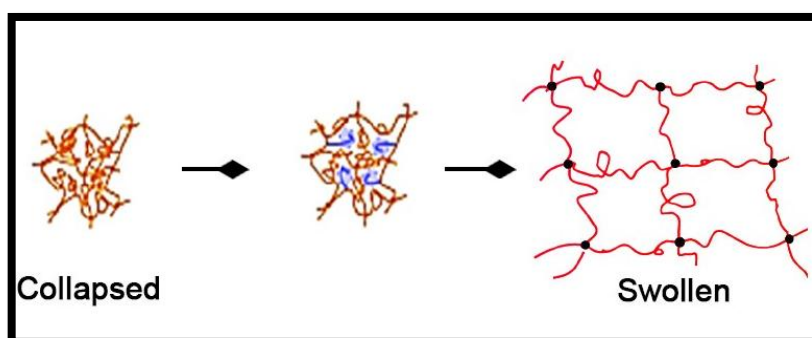


### 1.8.2. Swelling

Swelling occurs when the gelling agent interacts with the liquid, initiating solvation. A considerable amount of liquid is absorbed in response to this interaction, thus increasing the volume (Fig 1.4). The size of swelling is dependent on the force of attraction present in the molecular linkages of the gelling agent (Carter, 2000; Zatz and Kushla, 2005).

**Table 1.2.** Definition, advantages, and limitations of different types of pharmaceutical gels

Gel Type	Definition	Advantages	Limitations
<b>Niosomal or Proniosomal Gel</b>	Niosomal / proniosomal gels are simple gels containing Niosomes / proniosomes.	improved skin permeation with controlled release of the drug, improved stability chemically as well as physically, and are cost-effective in production	expensive, sensitive to moisture (hydrolysis), and quickly aggregate and sediment
<b>Emulgels</b>	Emulgels are gels in which water-in-oil or oil-in-water emulsions are added in a gel matrix.	Biofriendly, less greasy, greater spreadability, easy to remove, improved shelf-life, high patient acceptability due to the pleasant appearance, provides controlled release of drug, easy to prepare, cost-effective, improved thermodynamic stability, targeted drug delivery, and avoids of the first-pass effect	Skin irritation due to the presence of the surface-active agent(s)
<b>Bigels</b>	Bigels, as the name indicates, are a combination of hydrogel and oleogel (organogel), but they are free from any surfactant or stabilizer. They are prepared by mixing hydrogel and organogel at a high shear rate (rpm)	Easy to prepare, provide a synergistic effect, are free from surfactant; therefore, no skin irritation is reported due to surfactant, and more importantly, they can deliver both types of drug, i.e., lipophilic and hydrophilic	Phase separation
<b>Xerogels and Aerogels</b>	Xerogels are solid forms of a gel prepared by drying the wet silica gel at standard pressure with rapid shrinking of the gel, whereas in aerogels are compressible porous gel material from which liquid is removed conventionally.	More drug solubility leading to increased bioavailability, controlled release kinetics, Uniform drug distribution, easy incorporation of amorphous form, thermally stable	Silica aerogels are biodegradable
<b>Organogels</b>	Organogels are a network of a three-dimensional thermo-reversible gel consisting of an organogelator and a non-polar solvent	Thermoreversible, thermodynamically stable, biodegradable, biocompatible, and possess high transdermal permeability	Difficult to wash off
<b>Hydrogels</b>	hydrogels are also three-dimensional polymeric structures, but they contain a hydrophilic gelling agent (either synthetic or natural) that can hold a large amount of water	easy to prepare, cost-effective, biodegradable, adaptable to different compounds, and, it serves as a base for other advanced forms of gels such as liposomal gel, emulgels, microgels, nanogels, nanocomposite hydrogels, etc.	Microbial contamination



**Fig. 1.4.** Swelling of a gel.

### 1.8.3. Aging

When gelling agent forms a denser network with time, the phenomenon is known as aging (Carter, 2000; Zatz and Kushla, 2005).

### 1.8.4. Structure

The gelling agent forms a rigid network by interlinking its particles. The stress applied and the particles nature help in increasing the flow of gel (Patil et al., 2019).

### 1.8.5. Rheology

The gels obey non-Newtonian behavior as the solid dispersion in the gels follows pseudoplastic behavior, which indicates that the viscosity decreases as the shear rate increases (Patil et al., 2019).

## 1.9. APPLICATIONS OF GELS

Gel, a conventional dosage form, has various applications such as in cosmeceutical products (like shampoo, dentifrices, etc.), as catheter lubricant, as an adhering patch base, for electrocardiography suppository bases, etc. (Rathod and Mehta, 2015). Advancements in the field of the gels have remarkably increased the area of its application not only in the field of novel drug delivery systems (NDDS) but also for other purposes such as biomedical applications, hygiene (e.g., baby diaper, sanitary pads), in agriculture and horticulture, biotechnology, food packaging industry, cosmetic industry. Drug delivery through NDDS includes the controlled release of drugs, proteins, peptides, pesticides, nutrients, hormones, etc. However, drug delivery through the gel for any type of application requires the formation of gel that must be explicitly designed and must meet the desired properties (Ullah et al., 2015). Some of the examples of drugs recently incorporated into advanced gel formulations are listed in Table 1.3.

Gels, specifically hydrogels, in response to environmental changes, behave similarly to human organs, which direct their use towards medical implants, breast implants, artificial muscle diagnosis, bone implants, a decrease in thrombosis and intimal thickening in animals, etc. (Defail et al., 2006; Hill-West et al., 1994; Mendonça Munhoz et al., 2017). Thin films have also been prepared using organogels. They entrap the oil and hold it in a cross-linked network. This technique is used for a self-cleaning purpose and can also be used for different engineering metals (Liu et al., 2013).

Gels have a vast usage in sanitary pads to provide thinner pads with maximum absorbing power (Nagorski, 1994). The wastewater produced by the olive mill can be used as plant fertilizer, as hydrogel makes the wastewater immobilized, solving the polyphenolic issues (Davies et al., 2004).

**Table 1.3.** Example of drugs incorporated in different types of gels with their distinguish feature

Gel Type	Drug	Distinguishing Feature	Reference
Chitosan gel	Doxorubicin	Increased cytotoxicity	Taveira et al., 2009
Chitosan emulgel	Curcumin	Increased skin permeation	Thomas et al., 2017
Lecithin organogel	Sumatriptan succinate	Increased drug release from the skin	Agrawal et al., 2010
	Fenretinide	Improved stability and Shelf life	Esposito et al., 2013
	Crocin	Increased skin penetration	Esposito et al., 2016
	Metoprolol	Skin permeation enhancement	Varshosaz et al., 2013
Pluronic lecithin organogel (PLO)	Cyclobenzaprine	Skin permeation enhancement	Bryson et al., 2015
	Fluorescein	Increased transdermal permeation	Zhang et al., 2017
	Melatonin	Improved transdermal and transbuccal release	Flo et al., 2016
	Mefenamic acid	Topical controlled release	Jhawar et al., 2016
Bigel	Ciprofloxacin	Controlled drug delivery	Singh et al., 2014

	Vitamin E	Increased stability	Martinez et al., 2021
	Diclofenac	Improved drug delivery	Hamed et al., 2020
	diethylamine		
Hydrogel,	Ibuprofen	Improved rheological and	Hamed et al., 2018
oleogel, bigel		bioadhesive properties	
Alginate based	Ketoprofen,	Improved pharmacokinetic	Lovskaya and
aerogel	nimesulide,	properties	Menshutina, 2020
	loratidine		
Dextran and	5-Fluorouracil	Enzyme triggered targeted drug	Tiryaki et al., 2020
dextran		delivery to the colon	
aldehyde coated			
silica aerogel			
Silica aerogel	Celecoxib	Fast drug dissolution with	Jabbari-Gargari et
		controlled release	al., 2021
Silica xerogel	Nifedipine	Improved bioavailability	Zhang et al., 2020
Alginate-	Insulin	Improved wound healing	Rajalekshmy and
methacrylate			Rekha, 2021
xerogel			
Niosomal gel	Simvastatin	Enhanced anti-bacterial activity	Akbarzadeh et al.,
			2021
	Diclofenac	Improved therapeutic effect	Akbari et al., 2022
	sodium		
	Pilocarpine	Suitable for ocular drug delivery	Jain and Verma et
	hydrochloride		al., 2020
Proniosomal gel	Rutin	Improved cytotoxic effect on	Pinzaru et al., 2021
		melanoma cells	
	Naproxen	Possible alternate for oral	Shah et al., 2021
		naproxen	
	Ciclopirox	Prolonged drug release	Mahajan et al., 2021

Biodegradable hydrogels have been prepared and used as an alternative to polyacrylic hydrogel for personal and biomedical applications. Hydrogels formed by cross-linking CMC-Na or hydroxyethylcellulose with divinyl sulfone have been studied to treat edema as super absorbent (Sannino et al., 2003). In addition, hydrogels having distinctive mechanical properties have become the area of interest for drugs with a narrow absorption window (Omidian and Park, 2008; Omidian et al., 2005; Omidian et al., 2007).

Organogels have simplified the transdermal delivery of drugs. Lecithin organogels have been studied vastly for topical and transdermal drug delivery. Various compounds such as vitamins, hormones, peptides, amino acids, pain killers, anesthetics, and antifungal drugs have been studied for transdermal drug delivery and found successful for their delivery through lecithin organogels (Patel and Patel, 2021; Rajiv and Om, 2005; Sanapalli et al., 2018; Shaikh et al., 2006; Sushi et al., 2012; Willimann et al., 1992; Zhang et al., 2018). Fish oil has been used to prepare polymer-fish oil bigel and studied for transdermal drug delivery. The polymer-fish oil bigels showed excellent spreadability with improved skin permeation compared to the conventional hydrogel (Rehman et al., 2014). A list of pharmaceutical formulations of gels recently patented is given in Table 1.4.

All the types of gels discussed have several advantages with some limitations. The literature suggests that the hydrogels are most convenient to use because they are easy to prepare with an exceptional advantage that they can further be utilized to formulate other types of gels. This serves as the basis of developing NS based hydrogels.

**Table 1.4.** Recently patented gel formulations

S.No.	Patent No.	Formulation Title	Use	References
1.	US20210137101A1	Roach gel formulation	Insecticidal	Marwah et al., 2021
2.	WO2021034850A1	In-situ gel forming ophthalmic formulations containing difluprednate	Pain killer	Liang et al., 2021
3.	US10507264B1	Cross-linked hydrogel and method of making the same	Repairing, supplementing body tissue	Lowman et al., 2019
4.	US11026919B2	Formulation for soft anticholinergic analogs	Anticholinergic	Bodor et al., 2021
5.	US10561606B2	Injectable long-acting local anesthetic semisolid gel formulation	Controlled drug release	Shen and Gan, 2020
6.	US10881745B2	Formulation of nanostructured gels for increased agent loading and adhesion	Increased loading capacity	Van der Poll et al., 2021
7.	EP3351599B1	Corrosion-resistant adhesive sol-gel	Increased drug encapsulation	Schuette and Kinlen, 2019
8.	US10813882B1	In situ gelling formulation for reduced initial drug burst	Treating hyperlipidemia	Ahmed et al., 2020
9.	EP3372221B1	Hydrophobic gel based on vitamin E free from silicone products for topical application	Vitamin E free from silicone	Panin, 2019
10.	US11020410B2	Self-assembled gels formed with anti-retroviral drugs, prodrugs thereof, and pharmaceutical uses thereof	Treating HIV/hepatitis	Karp et al., 2021
11.	US10975217B2	Silica-based organogels via hexahydrotriazine-based reactions	Insulating materials	Boday et al., 2021
12.	EP3502779B1	Inherently photodegradable hydrogels or organogels for microfabrication	Photodegradable gel	Levkin and Li, 2020
13.	US10603406B2	Hydrogel for cell culture and biomedical applications	Cell culture and biomedical application	Huang, 2020
14.	US10500138B2	Polyphenols/PEG based hydrogel system for a dental varnish	Dental varnish	Jha and Simonton, 2019
15.	US10973956B2	Microporous hydrogel scaffolds for cell transplantation	Biomaterial implant (cell)	Kasputis et al., 2021

## REFERENCES

1. Agrawal, V., Gupta, V., Ramteke, S., Trivedi, P. (2010). Preparation and evaluation of tubular micelles of pluronic lecithin organogel for transdermal delivery of sumatriptan, *AAPS PharmSciTech*, 11:1718–1725.
2. Ahmed, T.A., Mussari, M.M., Omar, A.M., Khalid. M. (2020). In situ gelling formulation for reduced initial drug burst. Patent No. US10813882B1.
3. Akbari, J., Saeedi, M., Morteza-Semnani, K., Hashemi, S.M., Babaei, A., Eghbali, M., Mohammadi, M., Rostamkalaei, S.S., Asare-Addo, K., Nokhodchi, A. (2022). Innovative topical niosomal gel formulation containing diclofenac sodium (niofenac). *J Drug Target*, 30:108–117.
4. Akbarzadeh, I., Keramati, M., Azadi, A., Afzali, E., Shahbazi, R., Norouzian, D., Bakhshandeh, H. (2021). Optimization, physicochemical characterization, and antimicrobial activity of a novel simvastatin nano-niosomal gel against *E. coli* and *S. aureus*. *Chem Phys Lipids*, 234:105019.
5. Allan, S.H. (2012). Hydrogels for biomedical applications. *Adv Drug Deliv Rev*, 64:18–23.
6. Alnaief, M., Alzaitoun, M., Garcia Gonzalez, C.A., Smirnova, I. (2011). Preparation of biodegradable nanoporous microspherical aerogel based on alginate. *Carbohydr Polym*. 2011;84:1011–1018.
7. Amin, S., Rajabnezhad, S., Kohli, K. (2009). Hydrogels as potential drug delivery systems. *Sci Res Essay*, 30:1175–1183.
8. Andonova, V., Peneva, P., Georgiev, G.S., Toncheva, V.T., Apostolova, E., Peychev, Z., Dimitrova, S., Katsarova, M., Petrova, N., Kassarova, M. (2017). Ketoprofen-loaded polymer carriers in bigel formulation: An approach to enhancing drug photostability in topical application forms. *Int J Nanomedicine*, 12:6221–6238.
9. Attwood, D. (2002). Disperse systems. In: Aulton ME, Ed., *Pharmaceutics-The Science of Dosage Form Design*. London: Churchill Livingstone, UK, pp. 83–91, 528–529.
10. Balakrishnan, B., Jayakrishnan, A. (2015). Injectable hydrogels for biomedical applications. In: Nair LS, Ed., *Injectable Hydrogels for Regenerative Engineering*, 2nd ed., pp. 33–96.
11. Bhatia, R.B., Brinker, C.J., Gupta, A.K., Singh, A.K. (2000). Aqueous sol–gel process for protein encapsulation. *Chem Mater*, 12:2434–2441.
12. Biswal, S., Murthy, P., Sahu, J., Sahoo, P., Amir, F. (2008). Vesicles of non ionic surfactants (niosomes) and drug delivery potential. *Int J Pharm Sci Nanotech*, 1:1–8.
13. Boday, D.J., Garcia, J.M., Hedrick, J.L., Wertz, J.T., Wojtecki, R.J. (2021). Silica-based organogels via hexahydrotriazine-based reactions. Patent No. US10975217B2.
14. Bodor, N.S., Koleng, J.J., Angulo, D. (2021) Formulation for soft anticholinergic analogs. Patent No. US11026919B2.
15. Bryson, E., Hartman, R., Arnold, J., Gorman, G., Sweitzer, S., Asbill, S. (2015). Skin permeation and antinociception of compounded topical cyclobenzaprine hydrochloride formulations. *Int J Pharm Compd*. 2015;19:161–166.
16. Campoccia, D., Doherty, P., Radice, M., Brun, P., Abatangelo, G., Williams, D.F. (1998). Semisynthetic resorbable materials from hyaluronan esterification. *Biomater*, 19:2101–2127.
17. Carter, S.J. (2000) Cooper and Gunn's Tutorial Pharmacy, 6th ed., CBS Publishers and Distributors, New Delhi, India, pp. 68–72.
18. Charyulu, N.R., Joshi, P., Dubey, A., Shetty, A. (2021). Emulgel: A boon for enhanced topical drug delivery. *J Young Pharm*, 1:76.
19. Chellapa, P., Mohamed, A.T., Keleb, E.I., Elmahgoubi, A., Eid, A.M., Issa, Y.S., Elmarzughi, N.A. (2015) Nanoemulsion and nanoemulgel as a topical formulation. *IOSR J Pharm*, 5:43–47.
20. Cheng, Y., Lu, L., Zang, W., Shi, J., Cao, Y. (2012). Reinforce low density alginate based aerogels: preparation, hydrophobic modification and characterization. *Carbohydr Polym*, 88:1093–1099.

21. Choudhury, H., Gorain, B., Pandey, M., Chatterjee, L.A., Sengupta, P., Das, A., Molugulu, N., Kesharwani, P. (2017). Recent update on nanoemulgel as topical drug delivery system. *J Pharm Sci*, 106:1736–1751.
22. Co, E.D., Marangoni, A.G. (2018). Oleogels: An introduction. In: *Edible oleogels*, AOCS Press, Champaign, IL, USA, pp. 1–29.
23. Cuggino, J.C., Blanco, E.R., Gugliotta, L.M., Igarzabal, C.I., Calderón, M. (2019). Crossing biological barriers with nanogels to improve drug delivery performance. *J Controlled Release*, 307:221–246.
24. Davies, L.C., Novais, J.M., Martins-Dias, S. (2004). Detoxification of olive mill wastewater using superabsorbent polymers. *Environ Technol*, 25:89–100.
25. DeFail, A.J., Chu, C.R., Izzo, N., Marra, K.G. (2006). Controlled release of bioactive TGF- $\beta$ 1 from microspheres embedded within biodegradable hydrogels, *Biomater*, 27:1579–1585.
26. Dhawas, V., Dhabarde, D., Patil, S. (2020). Emulgel: A comprehensive review for novel topical drug delivery system. *Int J Recent Sci Res*, 11:38134–38138.
27. Di Michele, L., Fiocco, D., Varrato, F., Sastry, S., Eiser, E., Foffi, G. (2014). Aggregation dynamics, structure, and mechanical properties of bigels. *Soft Matter*, 10:3633–3648.
28. El-Laithy, H.M., Shoukry, O., Mahran, L.G. (2011). Novel sugar esters proniosomes for transdermal delivery of vinpocetine: preclinical and clinical studies. *Eur J Pharm Biopharm*, 77:43–55.
29. Esmaeely-Neisiany, R., Enayati, M.S., Sajkiewicz, P., Pahlevanneshan, Z., Ramakrishna, S. (2020). Insight into the current directions in functionalized nanocomposite hydrogels. *Front Mater*, 7:25.
30. Esposito, C.L., Kirilov, P., Roullin, V.G. (2018). Organogels, promising drug delivery systems: An update of state-of-the-art and recent applications. *J Control Release*, 271:1–20.
31. Esposito, E., Drechsler, M., Huang, N., Pavoni, G., Cortesi, R., Santonocito, D., Puglia, C. (2016). Ethosomes and organogels for cutaneous administration of crocin. *Biomed Microdev*, 18:1–12.
32. Esposito, E., Menegatti, E., Cortesi, R. (2013). Design and characterization of fenretinide containing organogels. *Mater Sci Eng: C*, 33:383–389.
33. Ferreira, S.A., Gama, F.M., Vilanova, M. (2013). Polymeric nanogels as vaccine delivery systems. *Nanomed: Nanotechnol Biol Med*, 9:159–173.
34. Flo, A., Calpena, A.C., Halbaut, L., Araya, E.I., Fernández, F., Clares, B. (2016). Melatonin delivery: transdermal and transbuccal evaluation in different vehicles. *Pharm Res*, 33:1615–1627.
35. Florence, A.T., Attwood, D. (2007). *FASTtrack: Physical Pharmacy*. Pharmaceutical Press, London, UK.
36. Ganesan, K., Budtova, T., Ratke, L., Gurikov, P., Baudron, V., Preibisch, I., Niemeyer, P., Smirnova, I., Milow, B. (2018). Review on the production of polysaccharide aerogel particles. *Materials*, 11:2144.
37. Gao, D., Xu, H., Philbert, M.A., Kopelman, R. (2008). Bioeliminable nanohydrogels for drug delivery. *Nano Lett*, 8:3320–3324.
38. Gonnet, M., Lethuaut, L., Boury, F. (2010). New trends in encapsulation of liposoluble vitamins. *J Control Release*, 146:276–290.
39. Goyal, S., Sharma, P., Ramchandani, U., Shrivastava, S.K., Dubey, P.K. (2011). Novel anti inflammatory topical gels. *Int J Pharm Biol Arch*, 2:1087–1094.
40. Guenther, U., Smirnova, I., Neubert, R.H.H. (2008). Hydrophilic Silica aerogels as dermal drug delivery systems- Dithranol as a model drug. *Eur J Pharm Biopharm*, 69:935–942.
41. Gupta, P.N., Mishra, V., Rawat, A., Dubey, P., Mahor, S., Jain, S., Chatterji, D.P., Vyas, S.P. (2005). Non-invasive vaccine delivery in transfersomes, niosomes and liposomes: a comparative study. *Int J Pharm*, 293:73–82.

42. Hajebi, S., Rabiee, N., Bagherzadeh, M., Ahmadi, S., Rabiee, M., Roghani-Mamaqani, H., Tahriri, M., Tayebi, L., Hamblin, M.R. (2019). Stimulus-responsive polymeric nanogels as smart drug delivery systems. *Acta Biomater*, 92:1–8.
43. Hamed, R., AbuRezeq, A.A., Tarawneh, O. (2018). Development of hydrogels, oleogels, and bigels as local drug delivery systems for periodontitis. *Drug Dev Ind Pharm*, 44:1488–1497.
44. Hamed, R., Mahmoud, N.N., Alnadi, S.H., Alkilani, A.Z., Hussein, G. (2020). Diclofenac diethylamine nanosystems-loaded bigels for topical delivery: Development, rheological characterization, and release studies. *Drug Dev Ind Pharm*, 46:1705–1715.
45. Harshitha, V., Swamy, M.V., Kumar, P., Rani, K.S., Trinath, A. (2020). Nanoemulgel: A process promising in drug delivery system. *Res J Pharm Dosage Forms Technol*. 2020;12:125–130.
46. Hill-West, J.L., Chowdhury, S.M., Slepian, M.J., Hubbell, J.A. (1994). Inhibition of thrombosis and intimal thickening by in situ photopolymerization of thin hydrogel barriers. *Proc Natl Acad Sci*, 91:5967–5971.
47. Huang, H. (2020). Hydrogel for cell culture and biomedical applications. Patent No. US10603406B2.
48. Ibrahim, A.A., Bosela, A.A., Ahmed, S.M., Mahrous, G.M. (2005). Proniosomes as a drug carrier for transdermal delivery of ketorolac. *Eur J Pharm Biopharm*, 59:485–490.
49. Ilomuanya, M.O., Hameedat, A.T., Akang, E.N., Ekama, S.O., Silva, B.O., Akanmu, A.S. (2020). Development and evaluation of mucoadhesive bigel containing tenofovir and maraviroc for HIV prophylaxis. *Future J Pharm Sci*, 6:1–2.
50. Jabbari-Gargari, A., Moghaddas, J., Hamishehkar, H., Jafarizadeh-Malmiri, H. (2021). Carboxylic acid decorated silica aerogel nanostructure as drug delivery carrier. *Microporous Mesoporous Mater*, 111220.
51. Jain, N., Verma, A. (2020). Preformulation studies of pilocarpine hydrochloride as niosomal gels for ocular drug delivery. *Asian J Pharm Clin Res*, 149–155.
52. Jain, N.K. (2006). *Pharmaceutical Product Development*. CBS Publishers & Distributors, New Delhi, India.
53. Jha, A., Simonton, T.C. (2019). Polyphenols/PEG based hydrogel system for a dental varnish. Patent No. US10500138B2.
54. Jhawar, V., Gupta, S., Saini, V. (2016). Formulation and evaluation of novel controlled release of topical pluronic lecithin organogel of mefenamic acid. *Drug Deliv*, 23:3573–3581.
55. Karg, M., Pich, A., Hellweg, T., Hoare, T., Lyon, L.A., Crassous, J.J., Suzuki, D., Gumerov, R.A., Schneider, S., Potemkin, I.I., Richtering, W. (2019). Nanogels and microgels: From model colloids to applications, recent developments, and future trends. *Langmuir*, 35:6231–6255.
56. Karp JM, Joshi N, Rioux D, Sherman NE, Pickering AJ, Gallin CF. Self-assembled gels formed with anti-retroviral drugs, prodrugs thereof, and pharmaceutical uses thereof. Patent No. US11020410B2, 2021.
57. Kashyap NK, Kumar N, Kumar MR. Hydrogels for pharmaceutical and biomedical applications. *Crit Rev Ther Drug Carrier Syst*. 2005;22:107–150.
58. Kasputis T, Skoumal M, Shea LD. Microporous hydrogel scaffolds for cell transplantation. Patent No. US10973956B2, 2021.
59. Khan S, Ullah A, Ullah K, Rehman NU. Insight into hydrogels. *Des Monomers Polym*. 2016;19:456–478.
60. Khandan, A., Jazayeri, H., Fahmy, M.D., Razavi, M. (2017). Hydrogels: Types, structure, properties, and applications. *Biomater Tissue Eng*, 4:143–169.
61. Koshani R, Tavakolian M, van de Ven TG. Natural emulgel from dialdehyde cellulose for lipophilic drug delivery. *ACS Sustain Chem Eng*. 2021;9:4487–4497.
62. Labarre, D., Ponchel, G., Vauthier, C. (2010). *Biomedical and Pharmaceutical Polymers*. Pharmaceutical Press, London, UK.

63. Larson, R.G. (1999). *The Structure and Rheology of Complex Fluids*, Oxford University Press, NY, USA.
64. Levkin, P., Li, L. (2020). Inherently photodegradable hydrogels or organogels for microfabrication. Patent No. EP3502779B1.
65. Liang, B., Zhang, M., Peng, H. 2021. In-situ gel forming ophthalmic formulations containing difluprednate. Patent No. WO2021034850A1.
66. Liu, H., Zhang, P., Liu, M., Wang, S., Jiang, L. (2013). Organogel-based thin films for self-cleaning on various surfaces. *Adv Mat*, 25:4477–4481.
67. Lovskaya, D., Menshutina, N. (2020). Alginate-based aerogel particles as drug delivery systems: Investigation of the supercritical adsorption and in vitro evaluations. *Mater*, 13:329.
68. Lowman, A., Brewer, E., Smith, N.G. (2019). Cross-linked hydrogels and method of making the same. Patent No. US10507264B1.
69. Lupi, F.R., Shakeel, A., Greco, V., Rossi, C.O., Baldino, N., Gabriele, D. (2016). A rheological and microstructural characterisation of bigels for cosmetic and pharmaceutical uses. *Mater Sci Eng: C*, 69:358–365.
70. Mahajan, S.S., Chaudhari, R.Y., Patil, V.R. (2021). Formulation and evaluation of topical proniosomal gel of ciclopirox for antifungal therapy. *Int J Pharm Investig*, 11:56–62.
71. Mahale, N.B., Thakkar, P.D., Mali, R.G., Walunj, D.R., Chaudhari, S.R. (2012). Niosomes: Novel sustained release nonionic stable vesicular systems—an overview. *Adv Colloid Interface Sci*, 183:46–54.
72. Marianecci, C., Carafa, M., Marzio, L.D., Rinaldi, F., Di Meo, C., Matricardi, P., Alhaique, F., Coviello, T. (2011). A new vesicle loaded hydrogel system suitable for topical applications: Preparation and Characterization. *J Pharm Pharm Sci*, 14:336–346.
73. Marianecci, C., Carafa, M., Marzio, L.D., Rinaldi, F., Di Meo, C., Matricardi, P., Alhaique, F., Coviello, T. (2011). A new vesicle loaded hydrogel system suitable for topical applications: Preparation and Characterization. *J Pharm Pharm Sci*, 14:336–346.
74. Martinez, R.M., Magalhães, W.V., da Silva Sufi, B., Padovani, G., Nazato, L.I., Velasco, M.V., da Silva Lannes, S.C., Baby, A.R. (2021). Vitamin E-loaded bigels and emulsions: Physicochemical characterization and potential biological application. *Colloid Surf B: Biointerfaces*, 201:111651.
75. Martins, A.J., Vicente, A.A., Pastrana, L.M., Cerqueira, M.A. (2020). Oleogels for development of health-promoting food products. *Food Sci Human Wellness*, 9:31–39.
76. Marwah, P., Tasz, M.K., Welzel, K. (2021). Roach gel formulations. Patent No. US20210137101A1.
77. Masotti, A., Vicennati, P., Alisi, A., Marianecci, C., Rinaldi, F., Carafa, M., Ortaggi, G. (2010). Novel Tween 20 derivatives enable the formation of efficient pH-sensitive drug delivery vehicles for human heptablastoma. *Bioorg Med Chem Lett*, 2:3021–3025.
78. Mendonça Munhoz, A., Santanelli di Pompeo, F., De Mezerville, R. (2017). Nanotechnology, nanosurfaces and silicone gel breast implants: current aspects. *Case Rep Plast Surg Hand Surg*, 4:99–113.
79. Murdan, S. (2005). Organogels in drug delivery. *Expert Opin Drug Deliv*, 2:489–505.
80. Nabi, S.A., Sheraz, M.A., Ahmed, S., Mustaan, N., Ahmad, I. (2016). Pharmaceutical gels: A review. *RADS J Pharm Pharm Sci*, 4:40–48.
81. Nagorski, H. (1994). Characterization of a new superabsorbent polymer generation. *Superabsorbent Polym*, 573:99–111.
82. Ojha, A., Ojha, M., Madhav, N.S. (2017). Recent advancement in emulgel: A novel approach for topical drug delivery. *Int J Adv Pharm*, 6:1–21.
83. Ollio, A., Ollio, J. (2009). The preparation of lignocellulosic aerogels from ionic liquid solutions. *Carbohydr Polym*, 75:125–129.
84. Omidian, H., Park, K., Rocca, J.G. (2007). Recent developments in superporous hydrogels. *J Pharm Pharmacol*, 59:317–327.



85. Omidian, H., Park, K. (2008). Swelling agents and devices in oral drug delivery. *J Drug Deliv Sci Technol*, 18:83–93.
86. Omidian, H., Rocca, J.G., Park, K. (2005). Advances in superporous hydrogels. *J Control Rel*, 102:3–12.
87. Panin, G. (2019). Hydrophobic gel based on vitamin e free from silicone products for topical application. Patent No. EP3372221B1.
88. Park, H., Park, K. (1996). Hydrogels in bioapplications, ACS Symposium Series, ACS Publications, NY, USA, pp. 2–10.
89. Patel, D., Patel, V. (2021). Development and characterization of pluronic lecithin organogel containing fluocinolone acetonide. *Drug Dev Ind Pharm*, 47:377–384.
90. Patil, P.B., Datir, S.K., Saudagar, R.B. (2019). A review on topical gels as drug delivery system. *J Drug Deliv Ther*, 9:989–994.
91. Pawbake, G.R., Shirolkar, S.V. (2020). Microemulgel: A promising approach to improve the therapeutic efficacy of drug. *J Crit Rev*, 7:1137–1143.
92. Peppas, N.A., Mikos, A.G. (2019). Preparation methods and structure of hydrogels. In: *Hydrogels in Medicine and Pharmacy* CRC press, Boca Raton, FL, USA, pp. 1–26.
93. Peppas, N.A., Barr-Howell, B.D. (2019). Characterization of the cross-linked structure of hydrogels. In: *Hydrogels in Medicine and Pharmacy*, CRC press, Boca Raton, FL, USA, pp. 27–56.
94. Pinelli, F., Perale, G., Rossi, F. (2020). Coating and functionalization strategies for nanogels and nanoparticles for selective drug delivery. *Gels*, 6:1–16.
95. Pinzaru, I., Tanase, A., Enatescu, V., Coricovac, D., Bociort, F., Marcovici, I., Watz, C., Vlaia, L., Soica, C., Dehelean, C. (2021). Proniosomal gel for topical delivery of rutin: Preparation, physicochemical characterization and in-vitro toxicological profile using 3D reconstructed human epidermis tissue and 2D Cells. *Antioxidants*, 10:85.
96. Puşcaş, A., Mureşan, V., Socaciu, C., Muste, S. (2020). Oleogels in food: A review of current and potential applications. *Foods*, 9:70.
97. Rajalekshmy, G.P., Rekha, M.R. (2021). Synthesis and evaluation of an alginate-methacrylate xerogel for insulin delivery towards wound healing applications. *Ther Deliv*, 12:215–234.
98. Rajiv, K., Om, P.K. (2005). Lecithin organogel as a potential phospholipidstructured system for topical drug delivery: A review. *AAPS PharmaSciTech*, 6:298–310.
99. Rathod, H.J., Mehta, D.P. (2015). A review on pharmaceutical gel. *Int J Pharm Sci*, 1:33–47.
100. Rehman, K., Amin, M.C., Zulfakar, M.H. (2014). Development and physical characterization of polymer-fish oil bigel (hydrogel/oleogel) system as a transdermal drug delivery vehicle. *J Oleo Sci*, 14:101.
101. Rehman, K., Zulfakar, M.H. (2014). Recent advances in gel technologies for topical and transdermal drug delivery. *Drug Dev Ind Pharm*, 40:433–440.
102. Sahoo, S., Kumar, N., Bhattacharya, C., Sagiri, S.S., Jain, K., Pal, K., Ray, S.S., Nayak, B. (2011). Organogels: Properties and applications in drug delivery. *Des Monomers Polym*, 14:95–108.
103. Salabat, A., Parsi, E. (2021). Ex vivo evaluation of celecoxib release from ionic liquid-based microemulsions and microemulgels for topical applications. *J Iran Chem Soc*, 18:1355–1361.
104. Sanapalli, B.K., Kannan, E., Balasubramanian, S., Natarajan, J., Baruah, U.K., Karri, V.V. (2018). Pluronic lecithin organogel of 5-aminosalicylic acid for wound healing. *Drug Dev Ind Pharm*, 44:1650–1658.
105. Sannino, A., Esposito, A., Rosa, A.D., Cozzolino, A., Ambrosio, L., Nicolais, L. (2003). Biomedical application of a superabsorbent hydrogel for body water elimination in the treatment of edemas. *Journal of Biomedical Materials Research Part A: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials*, 67:1016–10124.
106. Schuette, W.M., Kinlen, P.J. (2019). Corrosion resistant adhesive sol-gel. Patent No. US10508205B2.

107. Shah, H., Nair, A.B., Shah, J., Jacob, S., Bharadia, P., Haroun, M. (2021). Proniosomal vesicles as an effective strategy to optimize naproxen transdermal delivery. *J Drug Deliv Sci Technol*, 63:102479.
108. Shah, S., Rangaraj, N., Laxmikeshav, K., Sampathi, S. (2020). Nanogels as drug carriers—Introduction, chemical aspects, release mechanisms and potential applications. *Int J Pharm*, 581:119268.
109. Shaikh, I.M., Jadhav, K.R., Gide, P.S., Kadam, V.J., Pisal, S.S. (2006). Topical delivery of aceclofenac from lecithin organogels: Preformulation study. *Curr Drug Deliv*, 3:417–427.
110. Shakeel, A., Farooq, U., Iqbal, T., Yasin, S., Lupi, F.R., Gabriele, D. (2019). Key characteristics and modelling of bigels systems: A review. *Mater Sci Eng: C*, 97:932–953.
111. Shakeel, A., Lupi, F.R., Gabriele, D., Baldino, N., De Cindio, B. (2018). Bigels: A unique class of materials for drug delivery applications. *Soft Mater*, 16:77–93.
112. Shapiro, Y.E. (2011). Structure and dynamics of hydrogels and organogels: an NMR spectroscopy approach. *Prog Polym Sci*, 36:1184–1253.
113. Shehata, T.M., Ibrahim, M.M., Elsewedy, H.S. (2021). Curcumin niosomes prepared from proniosomal gels: In vitro skin permeability, kinetic and in vivo studies. *Polym*, 13:791.
114. Shen, H.R., Gan, N. (2020). Injectable long-acting local anesthetic semi-solid gel formulations. Patent No. US10561606B2.
115. Singh, V.K., Anis, A., Al-Zahrani, S., Pradhan, D.K., Pal, K. (2014). Molecular and electrochemical impedance spectroscopic characterization of the carbopol based bigel and its application in iontophoretic delivery of antimicrobials. *Int J Electrochem Sci*, 9:5049–5060.
116. Soni, G., Yadav, K.S. (2016). Nanogels as potential nanomedicine carrier for treatment of cancer: A mini review of the state of the art. *Saudi Pharmaceut J*, 24:133–139.
117. Soni, K.S., Desale, S.S., Bronich, T.K. (2016). Nanogels: An overview of properties, biomedical applications and obstacles to clinical translation. *J Control Rel*, 240:109–126.
118. Sreekumar, M., Mathan, S., Mathew, S.S., Dharan, S.S. (2020). Bigels: An Updated Review. *J Pharm Sci Res*, 12:1306–1308.
119. Stergar, J., Maver, U. (2016). Review of aerogel-based materials in biomedical applications. *J Sol-Gel Sci Technol*, 77:738–752.
120. Sun, Z., Li, Z., Qu, K., Zhang, Z., Niu, Y., Xu, W., Ren, C. (2021). A review on recent advances in gel adhesion and their potential applications. *J Mol Liq*, 115254.
121. Sushi, I.R., Santosh, S.B., Vaibhav, U., Mishra, V., Gahane, A., Jain, S.K. (2012). Lecithin organogel: A unique micellar system for the delivery of bioactive agents in the treatment of skin aging. *Acta Pharmaceut Sinica B*, 2:8–15.
122. Taveira, S.F., Nomizo, A., Lopez, R.F. (2009). Effect of the iontophoresis of a chitosan gel on doxorubicin skin penetration and cytotoxicity. *J Control Release*, 134:35–40.
123. Thomas, L., Zakir, F., Mirza, M.A., Anwer, M.K., Ahmad, F.J., Iqbal, Z. (2017). Development of curcumin loaded chitosan polymer based nanoemulsion gel: In vitro, ex vivo evaluation and in vivo wound healing studies. *Int J Biol Macromol*, 101:569–579.
124. Tiryaki, E., Elalmış, Y.B., İkizler, B.K., Yücel, S. (2020). Novel organic/inorganic hybrid nanoparticles as enzyme-triggered drug delivery systems: Dextran and dextran aldehyde coated silica aerogels. *J Drug Deliv Sci Technol*, 56:101517.
125. Ullah, F., Othman, M.B., Javed, F., Ahmad, Z., Akil, H.M. (2015). Classification, processing and application of hydrogels: A review. *Mater Sci Eng: C*, 57:414–433.
126. United States Pharmacopeia 30/National Formulary 25, United States Pharmacopeial Convention, Inc., Rockville, MD, 2021; Electronic version.
127. Uros, M., Aliaz, G., Marian, B., Odon, P. (2007). Novel hybrid silica xerogels for stabilization and controlled release of drug. *Int J Pharm*, 330:164–174.
128. van der Poll, D.G., Blasioli, D.J., Zugates, G.T. (2021). Formulation of nanostructured gels for increased agent loading and adhesion. Patent No. US10881745B2.

129. Varshosaz, J., Andalib, S., Tabbakhian, M., Ebrahimzadeh, N. (2013). Development of lecithin nanoemulsion based organogels for permeation enhancement of metoprolol through rat skin. *J Nanomat*, 2013.
130. Vashist, A., Vashist, A., Gupta, Y.K., Ahmad, S. (2014). Recent advances in hydrogel based drug delivery systems for the human body. *J Mat Chem B*, 2:147–166.
131. Verma, A., Tiwari, A., Saraf, S., Panda, P.K., Jain, A., Jain, S.K. (2020). Emerging potential of niosomes in ocular delivery. *Expert Opin Drug Deliv*, 1–7.
132. Vintiloiu, A., Leroux, J.C. (2008). Organogels and their use in drug delivery—a review. *J Control Release*, 125:179–192.
133. Willmann, H., Walde, P., Luisi, L., Gazzaniga, A., Stroppolo, F. (1992). Lecithin organogels as matrix for transdermal transport drugs. *J Pharm Sci*, 81:871–874.
134. Zatz, J.L., Kushla, G.P. (1989). Gels, In: Lieberman HA., Rieger MM, Banker GS. *Pharmaceutical Dosage Form: Disperse Systems*, 2nd ed., Vol. 2, Marcel Dekker, NY, USA, pp. 399–421.
135. Zhang, P., Jiang, Q., Zheng, Y., Li, J. (2020). Double-nano silica xerogel contributes to establish nifedipine delivery system with superior delivery effect. *Microporous Mesoporous Mater*, 296:109996.
136. Zhang, Q., Song, Y., Page, S.W., Garg, S. (2018). Evaluation of transdermal drug permeation as modulated by lipoderm and pluronic lecithin organogel. *J Pharm Sci*, 107:587–594.
137. Zhang, Y., Gao, W., Chen, Y., Escajadillo, T., Ungerleider, J., Fang, R.H., Christman, K., Nizet, V., Zhang, L. (2017). Self-assembled colloidal gel using cell membrane-coated nanosponges as building blocks. *ACS Nano*, 11:11923–11930.