



## PREVALENCE OF ENDOGLOBULAR HEMOTROPIC PARASITES IN OPC SHEEP IN THE MUNICIPALITY OF SINCELEJO AND SAVANNA SUB-REGION, DEPARTMENT OF SUCRE-COLOMBIA

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### ABSTRACT

Infection caused by hemoparasite is one of the most prevalent diseases in tropical and subtropical countries in the world. Anaplasmosis and babesiosis are diseases caused by intracellular hematopoietic agents that are part of the parasitic complex. Hematotropic agents (*Anaplasma ovis*, *Anaplasma marginale*, *Trypanosoma vivax*, *Trypanosoma melophagium*, *Babesia ovis* and *Babesia motasi*) are common causes of hematopoietic infections in sheep. Thus, the aim of the present study was to determine the frequency of endoglobular hemotropic parasites in Colombian hair sheep (OPC) in municipality of Sincelejo and Savanna Sub-region of the department Sucre- Colombia. A cross-sectional study was carried out at convenience, according to availability in each farm during the years 2020-2022, totaling 400 animals. The capillary microcentrifuge technique was used, where the percentage volume of red blood cells (Haematocrit) was determined by reading on a Hawksley microhaematocrit reader table, then blood smears were made, stained with WRIGTH dye and evaluated under a light microscope. Incident variables in the parasitaemia of the animals were considered, such as sex, location and Body Condition Score, looking for a relationship with the parasite positivity that the animals could present. The results of the analysis showed 165 positive samples, representing a prevalence for *Anaplasma* spp of 14.26%; *Babesia* spp 14.5% and for both agents 12.5% in OPC. There was a significant relationship ( $p < 0.05$ ) between animal origin, gender (Male-Female) and Body Condition Score, with the degree of prevalence that the animals possessed. In conclusion, *Anaplasma* spp and *Babesia* spp, is present in the municipality of Sincelejo and Savanna Sub-region (Sucre- Colombia), and these can occur in OPC animals from different localities, sex and Body Condition Score.

**Key words:** hemoparasite, sheep, *Babesia* spp, *Anaplasma* spp, endoglobular.

## INTRODUCTION

In Colombia there is a diversity of sheep, exploited under a traditional production scheme, the largest population of animals correspond to the short-haired breed, known as Camuras, currently OPC, highly adapted to diverse agro-climatic conditions (Flórez et al., 2020; Noriega et al., 2022). Sheep production in Colombia, especially on the north coast, is developed by small producers, basing its development specifically on the OPC breed, which has a series of limitations for optimal performance (infrastructure, reproductive, nutritional, genetic and sanitary management), reflected in low production indices (Hernández et al., 2023). Studies by Al Kalaldehy et al. (2019), showed that parasite infections were one of the most important problems in sheep and goats, causing severe production constraints in small ruminants, especially those raised by marginal producers in a low external input system. In this sense, infections caused by hemoparasites have a high rate of occurrence in tropical and subtropical countries (Torres et al., 2021; Nyifi et al., 2023; Torres et al., 2023), with the highest incidence in animal production systems being caused by Babesiosis (*Babesia* spp), Anaplasmosis (*Anaplasma* spp) and Trypanosomiasis (*Trypanosoma* spp), causing a considerable reduction in meat and milk production in livestock (Avila et al., 2013, Torre et al., 2021).

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Haematopoietic agents (*Anaplasma ovis*, *Anaplasma marginale*, *Trypanosoma vivax*, *Trypanosoma melophagium*, *Babesia ovis* and *Babesia motasi*), are a common cause of blood borne infections in sheep, Avila et al. (2013) reported the presence of *Anaplasma* spp. in 73.7%, in a sheep population, studied in the department of Antioquia-Colombia. The identification of haemoparasitic disease is of great help and allows better targeting of intervention (Plaza et al., 2019). Haemoparasitic disease is suspected when there is fever, accompanied by anaemia, animal death and/or abortion, presence of oedema, haemoglobinuria or jaundice. The department of Sucre (Colombia) is one of the thirty-two departments of Colombia, located in the northern part of the country. It is made up of 26 municipalities and 275 townships, which, taking into account the criteria of geographical location, political-administrative division, economic vocation, inter-municipal relations, historical and socio-cultural links, are grouped into five physiographic Sub-regions: Gulf of Morrosquillo, Mountains of María, Savannas, San Jorge and La Mojana, the majority of the population lives in rural areas and depends on agriculture as the main economic activity. Thus, the objective of the present study was to determine the frequency of endoglobular hemotropic parasites in OPC sheep from the municipality of Sincelejo and Savannas Sub-region (department of Sucre, Colombia).

## 2. MATERIALS AND METHODS

**2.1 Type of study.** A descriptive cross-sectional study was conducted on OPC animals at convenience, according to availability on each farm during the years 2020-2022 (Manterola & Otzen, 2014).

**2.2 location.** OPC sheep farms in the Savannas Sub-region of the department of Sucre (Municipalities of Sincé, El Roble, San Pedro, Sampués, Los Palmitos, Galeras, Buenavista, Corozal, San Juan de Betulia) and Sincelejo, Colombia (Figure 1). The life zone of the study site is classified as tropical dry forest (bs-t), with temperatures ranging between 25.5 and 28.7°C, precipitation fluctuating between 990 and 1275 mm per year, relative humidity of 80% (Montes et al., 2022).

**2.3 Sample size and study population.** Considering that municipality of Sincelejo and the Savannas Sub-region have a sheep census to 2023 (ICA-2023) of 12,151 head, a sample size of 400 animals was calculated (confidence level of 95%;  $\alpha= 0.05$ ;  $Z= 1.96$  and a margin of error of 5%), distributed

in municipality of Sincelejo and the 10 municipalities that make up the Savannas Sub-region. A stratified random sampling was carried out, to take 40 samples per municipality, for a total of 400 animals. The farms per municipality (4 in total) were subject to selection by convenience, taking into account the geographical distribution of the municipality, and 10 animals were taken at random from each farm. The selected production units usually carried out traditional management for the breeding and exploitation of sheep.

**2.4 Inclusion and exclusion criteria.** The animals were sampled from three months of age, between males and females, no younger animals were considered, due to maternal acquired immunity, the selected animals were numbered with Ear Tags Numbered.

## 2.5 Data collection techniques and instruments.

**2.5.1 Blood sampling and diagnosis.** Blood samples were collected by venipuncture of the jugular vein. Then, 5 ml of peripheral blood was extracted in a vacuum vacutainer tube without anticoagulant and with anticoagulant (EDTA), duly identified with the number of the specimen sampled, kept refrigerated, and then processed in the laboratory.



**Figure 1.** Study area: municipality of Sincelejo and Savanna Sub-region (Department of Sucre, Colombia). Source: Taken from Montes et al., (2022).

**2.5.2 Data processing and analysis.** The capillary microcentrifugation technique (TMS, described by Woo 1969) was used, where the percentage volume of red blood cells (haematocrit) was determined by reading on a Hawksley micro-haematocrit reader (Dill and Cost, 1974). Afterwards, a blood smear and staining with wright dye was performed, and evaluated under a light microscope

with an immersion objective, according to the procedure, which allows the evaluation of intracellular parasitic forms morphologically compatible with *Anaplasma* spp and *Babesia* spp (López et al., 2014; Calderón et al., 2016).

With the results obtained, a database was elaborated with the evaluated variables where sheep without haematozoa and with haematozoa were compared; where the latter category was established with at least the diagnosis of only one group. The comparison of the groups was implemented with the t-student test for quantitative variables; all these calculations were carried out using the statistical software R. 4.3.02.

Equation 1 was applied to find the incidence of the etiological agent of endoglobular hemotropic parasite diseases.

$$\text{Equation 1.} = \frac{\text{Number of endoglobularhemotropic positive animals}}{\text{Total animals sampled}} \times 100$$

Equation 2 was applied to relate the prevalence of endoglobular hemotropic parasites with the variable Body Condition Score (BCS) in OPC sheep from the municipality of Sincelejo and Savanna Sub-region (Sucre – Colombia).

$$\text{Equation 2.} = \frac{\text{Number of positive animals with BCS (1 – 5)}}{\text{Total animals sampled}} \times 100$$

BCS was determined by palpation of the lumbar region based on a scale of 1-5, (with an increment of 0.5), where 1: very thin and 5: obese (Russel et al., 1969; Canul et al., 2022), with a score between 3 and 4 being considered optimal. The qualitative BCS classification would be in the order of: obese (> 4.0), fat (3.5-4.0), normal (3.0-3.5), thin (2.5 - 3.0), very thin (< 2.5).

To determine the prevalence ratio of endoglobular hemotropic positive animals to haematocrit, equation 3 was implemented for the resulting

$$\text{Equation 3.} = \frac{\text{Haematocrit of endoglobular hemotropic parasite – positive animals}}{\text{Total animals sampled}} \times 100$$

For ease of analysis the haematocrit obtained were classified into ranges for tabulation. The ranges used were as follows: Haematocrit ranging from 20 to 30, from 31 to 40, from 41 to 50 and haematocrit greater than 50. The animals were divided into those positive for haematopoietic parasites, either *Anaplasma* spp, *Babesia* spp or both, or those negative for these parasites. The ocular mucosa was evaluated according to the color of the mucosa, taking into account the following classification: Congestive; Sub-Icteric; Pale; Normal.

**2.4.3 Survey of farm owners.** In each sampled herd, the respective owners were surveyed in order to obtain animal information such as age (category), sex, zoning and presence of ticks, this information was used in the data analysis. The surveys were open-ended and the variables to be considered were nominal and binary.

### 3. RESULTS AND DISCUSSION

Taking into account the main objective of the present work, the frequency of endoglobular hemotropic in OPC sheep in the municipality of Sincelejo and Savanna Sub-region, department of Sucre, was 41.25% (165/400), being the main pathogen association *Anaplasma* spp. and *Babesia* spp. or co-infected by both (Table 1).

**Table 1.** Overall prevalence of endoglobular hemotropic in OPC sheep

Specie	N° of samples	Positives		Negatives		I.C.95%
		N°	%	N°	%	
Ovis aries (OPC)	400	165	41.25	235	58.75	0.024

Table 2 presents the overall infestation rates by species of endoglobular hemotropic, of which 14.26% (57/400) were positive for *Anaplasma* spp, 14.5% (58/400) were positive for *Babesia* spp, and 12.5% (50/400) were positive for both agents (co-infection). The possible reasons for the high number of parasitized animals observed in the study may be due to the poor parasite control carried out on the farms, sometimes these animals are bathed for tick control.

The results found in the present study differ from those reported by Martinez and Tatis (2001) in the department of Sucre - Colombia, where the frequencies of infection by *Anaplasma* spp., *Babesia* spp. and *Trypanosoma* sp. were 90%, 0% and 0% respectively. Likewise, Li et al (2020) reported prevalence of *Babesia* spp and *Anaplasma* spp (67.7%) in sheep from border regions of northwest China. In general, different prevalence values are reported in the literature (Rahravani et al., 2023; Arif et al., 2023; Taqadus et al., 2023; Onyiche & MacLeod, 2023), which may be attributed to the difference in micro and macro climate of each region, tick species, tick habitat and landscape between regions.

**Table 2.** Prevalence of endoglobular hemotropic species found in sheep OPC

N° OPC	Positives		Positives		Positives	
	Anaplasma spp.	%	Babesia spp.	%	y Babesia spp.	%
400	57	14.26	58	14.5	50	12.5

A good indicator of the general state of the animals is the BCS, although it is not a specific and strict parameter in its measurement, it is of great help in the clinical and productive evaluation of an animal. In Colombia, livestock regions located in tropical areas are considered enzootic for the haemoparasites (González et al., 2014), animals become reservoirs, so it is necessary to have diagnostic techniques that allow the detection of carrier animals, in order to know the prevalence of the disease in the regions (Jaimes et al., 2017; Vargas et al., 2019).

Table 3 describes the relationship of OPC sheep diagnosed positive for endoglobular hemotropic parasites with respect to BCS. Of the 57 animals sampled for *Anaplasma* spp, 78.3% (45/57), presented a very thin BCS (< 2.5), 12. 3% (7/56) had a thin BCS and only 7% (4/57) of the animals had a normal BCS.

Of the 58 animals positive for *Babesia* spp, just over 90% of them had a poor BCS (Table 3), which can be attributed to the inappetence experienced by animals with this disease due to the secretion of some substances that disrupt the feeding process, In addition, more than 96% of the total number of animals in the study that tested positive for both agents had a BCS score below normal (3.0-3.5), resulting in low productive animals (Chochlakis, et al., 2009; Bauer et al.,2021; Onyiche et al., 2022; Onyiche & MacLeod, 2023). A significant difference ( $p= 0.039$ ), for *Anaplasma* spp; *Babesia* spp ( $p= 0.029$ ) and *Anaplasma* spp and *Babesia* spp ( $p= 0.042$ ), was observed in the very thin BCS category (< 2.5) with respect to the other categories. In contrast, Mannat et al. (2023) reported a non-significant effect of BCS on animals diagnosed positive for endoglobular hemotropic parasites.

**Table 3.** Ratio of OPC sheep diagnosed positive for endoglobular hemotropic parasites to BCS.

BCS	Positives		Positives		Positives	
	Anaplasma spp.	%	Babesia spp.	%	y Babesia spp.	%
Obese > 4.0	0	0.0	0	0	0	0

Fat (3.5-4.0)	1	1.8	1	1.7	0	0
Normal (3.0-3,5)	4	7.1	3	5.2	2	4
Slim (2.5 - 3.0)	7	12.5	6	10.3	5	10
Very thin< 2.5*	44	78.6	48	82.8	43	86
Total	57	100	58	100	50	100

\*Significant differences (p < 0.05)

The results obtained evaluating the sex variable (Table 4), showed a significant difference (p= 0.036) of the sex in the OPC sheep, with respect to the prevalence of endoglobular hemotropic parasites, females have the highest prevalence 92. 7% (153/165), compared to males 7.3% (12/165), there is a relationship between the sex of the animal and the presence of endoglobular hemotropic parasites in OPC sheep, these results are concordant with those presented by Abdelsalam et al, (2023) and Eisawi et al., (2020). On the contrary, some results found in the literature state that the prevalence of endoglobular hemotropic parasites in sheep is not affected by gender (Naeem et al., 2023; Tamrat et al., 2023; Hamzah & Hasso,2019; Egbe-Nwiyi et al., 2018; Shah et al., 2017).

**Table 4.** Sex-specific prevalence of endoglobular hemotropic parasites in OPC sheep.

Sexo	N°	Positivos		Negativos	
		N°	%	N°	%
Males	82	12	7.3	70	29.8
Females*	318	153	92.7	165	70.2
Total	400	165	100	235	100

\*Significant differences (p < 0.05)

The origin of the animal was evaluated with the possibility of zoning the distribution of the parasite in the localities of municipality of Sincelejo and Savanna Sub-region (Sucre-Colombia), finding significant differences (p= 0.042). The municipalities with the highest prevalence of endoglobular hemotropic parasites were Galera, San Pedro, Sampués, Sincelejo and Los Palmito with an overall prevalence equal to or higher than 45% (Table 5). In these municipalities, the presence and management of OPCs are closely related to cattle production, where animals tend to become infected with the pathological agent at an early age, allowing them to reach certain levels of coexistence between host and parasite. Noaman and Alireza, (2023), reported significant differences in parasite prevalence between the different geographical areas of their study (De la Fuente et al., 2007; Rahravani et al., 2023; Prajapati et al., 2023).

**Table 5.** Prevalence of endoglobular hemotropic parasites with respect to zoning by municipality in OPC sheep.

Municipality	N° de sites	Sample	Positive	%	Negative	%
Sincé	4	40	12	30.0	28	70.0
El Roble	4	40	16	40.0	24	60.0
San Pedro*	4	40	18	45.0	22	55.0
Sampués*	4	40	18	45.0	22	55.0
Los Palmitos*	4	40	19	47.5	21	52.5
Galeras*	4	40	22	55.0	18	45.0
Buena Vista	4	40	12	30.0	28	70.0
Corozal	4	40	16	40.0	24	60.0
San Juan de Betulia	4	40	15	37.5	25	62.5
Sincelejo*	4	40	17	42.5	23	57.5
Total	40	400	165		235	

\*Significant differences (p < 0.05)

The low prevalence in some localities (Table 5) may be attributed to frequent parasite control plans against ticks, which compensates for the decrease in the vector population. Similarly, the type of vegetation, landform, sampling areas and possibly the type of animal management may also influence the degree of prevalence of endoglobular hemotropic parasites. A statistically significant relationship

between parasite treatments has been documented in the literature, favoring the acquisition of protective immunity from an early age and the absence of clinical signs in the animals over time (Reátegui et al., 2023).

Table 6 describes the haematocrit concentration in OPC sheep positive for *Babesia* spp and *Anaplasma* spp. It can be observed that 61.4% (35/57) of the animals positive for *Anaplasma* spp. had a haematocrit between 20 and 30, possibly due to intrasplenic haemolysis by the endothelial reticulum system, produced by this type of bacteria. The 28% (16/57) of the animals positive to *Anaplasma* spp, presented haematocrit values between 31 and 40, and only 10.6% of these animals presented haematocrit in the range between 41 and 50, given the above, it is presumable that this intra-erythrocytic bacterium is the direct cause of anaemia in these animals. According to the literature, a goat or sheep infected with *Anaplasma* spp. that presents a haematocrit higher than 25% is considered an asymptomatic carrier, a condition commonly found in goats and sheep with anaplasmosis; which is also considered an epidemiological condition that makes the animals a reservoir for the microorganism (Avila et al, 2013; Arece et al., 2015), studies developed by Rahravani et al., (2023) and Nangru et al., (2023), found a statistically significant association between semi-yellow conjunctiva, mean red blood cell count, mean corpuscular haemoglobin and haemoglobin concentration in all haemoparasites infections.

**Table 6.** Haematocrit concentrations in *Babesia* spp. and *Anaplasma* spp. positive OPC sheep in OPC sheep.

Haematocrit	Positive <i>Anaplasma</i> spp		Positive <i>Babesia</i> spp		Positive <i>Anaplasma</i> spp y <i>Babesia</i> spp	
	Count	%	Count	%	Count	%
20-30	35	61.4	48	82.8	43	86.0
31-40	16	28.0	8	13.8	6	12.0
41-50	6	10.6	2	3.4	1	2.0
>50	0	0.0	0	0.0	0	0.0
Total	57	100	58	100	50	100

Regarding the animals positive for *Babesia* spp, 82. This is attributable to the physiopathogenesis of *Babesia* spp, which apart from directly producing lysis on the red blood cell, also causes the immune system to destroy other red blood cells, resulting in severe anaemia. 13.8% of the OPCs positive for *Babesia* spp, had haematocrit values between 31 and 40, and only 3.4% were in the range between 41 and 50. The animals parasitized by this agent were the ones that showed the most sanitary deterioration, in relation to the animals parasitized by both agents, 86 % of the OPC sheep presented haematocrit values between 20 and 30, so it can be said that the *Babesia*-haematocrit ratio predominated over the *Anaplasma*-haematocrit ratio. Similarly, it was found that 12 % of these OPC sheep had values in the range between 31 and 40. It should be noted that the haematocrit is the most important measure to determine the degree of anaemia of an animal, and one of the most important clinical signs of the diseases produced by these microorganisms, therefore, it is essential to know the haematocrit-haematopoietic agent ratio (Nyifi & Bilbonga, 2023).

Table 7 describes the relationship between the color of the mucosal membranes in OPC sheep with the presence of *Babesia* spp and *Anaplasma* spp. 88 % of the 57 OPC sheep that were positive for *Anaplasma* spp, presented pale mucous membranes, which is explained by the type of haemolysis that this produces, being intrasplenic (extravascular) does not increase the production of bile pigments by the liver, as a consequence the membranes turn to a whitish color, 12 (7/57)% of this group, presented sub-icteric coloring, which can be attributable to some other type of hepatic problem or to the presence of *Anaplasma* spp, without being observed in the samples, since the technique used in this study is dependent on the expertise of the laboratory technician.

**Table 7.** Mucosal color of OPC sheep sampled during the study.

Hematocritos	Positive Anaplasma spp		Positive Babesia spp		Positive Anaplasma spp y Babesia spp	
	Anaplasma spp	%	Babesia spp	%	spp y Babesia spp	%
Congestive	0	0.0	20	34.5	11	22.0
Sub- Icteric	7	12	37	63.8	37	74.0
Pale	50	88	1	1.7	2	4.0
Normal	0	0.0	0	0.0	0	0.0
Total	57	100	58	100	50	100

Of the group of animals positive to Babesia spp, 63.8% (37/58) presented sub-icteric mucosa, possibly this could be caused by the intravascular haemolysis produced by Babesia spp, the free Heme group in blood is converted into biliverdin by the SRE, and then conjugated in the liver, to bilirubin diglucuronide and its high levels cause yellowish pigmentation of the mucosa, the remaining 34. The remaining 34.5% (20/58) of this group had congestive mucous membranes, which could be related to dehydration of the animals. Of the 50 animals that were positive for both agents, 22% (11/50) had congestive mucosa, and 74% (37/50) had sub-icteric mucosa, indicating a dominance of clinical signs associated with Babesia over clinical signs of Anaplasma. Several authors have related the conjunctival coloring of the eye with the presence of anaemia, finding association with parasite loads in animals (B. Adehanom et al., 2015; Adamu et al., 2020; Şahin et al.,2021; Coello-Peralta et al., 2022; Rahravani et al., 2023).

Table 8 shows the infestation level of OPC sheep positive for endoglobular hemotropic, none of the OPC evaluated presented an infestation level of 0.07%. Despite the high frequency of infection by Anaplasma spp and Babesia spp, no symptomatic animals were observed, which may be related to the low infestation levels (0.01) found in the positive animals. The samples tested showed an average infection and infestation percentage of 0.014% for Anaplasma spp; 0.017% for Babesia spp and 0.014% for Anaplasma spp and Babesia spp respectively.

**Table 8:** Infestation level (%) of endoglobular hemotropic positive OPC sheep.

Hemotropic	Level of infestation (%)							
	0.01	0.02	0.03	0.04	0.05	0.06	0.07	Total
Anap	44	7	3	1	2	0	0	57
Bab	42	4	4	4	3	1	0	58
Anap y Bab	40	2	1	3	3	1	0	50

Anap= Anaplasma spp; Bab =Babesia spp; Anap y Bab = Anaplasma spp y Babesia spp

#### 4. CONCLUSIONS

Anaplasma spp and Babesia spp are present in the municipality of Sincelejo and the Savanna Sub-region (Sucre-Colombia), and can occur in OPC sheep of different localities, sex and Body Condition Score. The agro-ecological conditions of the study area are conducive to the development of specific vectors of these microorganisms, so it is appropriate to implement prophylactic measures to reduce the incidence of these agents, because they are directly related to economic losses in the sheep population worldwide, significantly deteriorating the health status of animals and in some cases causing death.

#### 5. CONFLICT OF INTEREST

There is no conflict of interest related to the subject matter of the work.

#### 6. CONTRIBUTION OF THE

Conceptualization, data curation, formal analysis, research, methodology, supervision, validation, writing - original draft, writing - revision and editing: Alexander Pérez C, Donicer Montes V and Gabriela Flórez M.



## REFERENCES

1. Abdelsalam, MA., Felefel, W., Fadl, S. et al. (2023). Molecular prevalence and associated infection risk factors of tick-borne protozoan and rickettsial blood pathogens in small ruminants. **BMC Vet Res** 19, 138 <https://doi.org/10.1186/s12917-023-03702-4>
2. Adamu, M., Dzever, P., Ikurior, S. et al. (2020). Validation of the FAMACHA© system for detecting anaemia and helminthosis in West African Dwarf sheep in Makurdi, Benue State, Nigeria. **Comp Clin Pathol** 29:965–970). <https://doi.org/10.1007/s00580-020-03157-1>
3. Al Kalaldeh M., Gibson J., Duijvesteijn N., Daetwyler H., MacLeod I., Moghaddar N., et al. (2019). Using imputed whole-genome sequence data to improve the accuracy of genomic prediction for parasite resistance in Australian sheep **Genet Sel Evol**,51(1):32. <https://doi.org/10.1186/s12711-019-0476-46>.
4. Arece García, Javier., Sanabria, Argemiro., Soca, Mildrey, da Fonseca., Adivaldo H., Fidlarczyk Maciel., Raisa, da Silva., Larissa Clara., Tomaz, Aline Maria., & Zen Gianfrancisco, Olivia. (2015). Relación de algunos indicadores sanguíneos con la infestación de parásitos gastrointestinales en ovinos. **Revista de Salud Animal**, 37(2), 133-135.
5. Arif, M., Saeed, S., Bashir, A., Farooq, M., Nasreen, N., Khan, A., & Chen, C. C. (2023). Molecular prevalence and phylogeny of *Anaplasma marginale*, *Anaplasma ovis* and *Theileria ovis* in goats and sheep enrolled from a hill station in Punjab, Pakistan. **Plos one**, 18(11), e0291302.
6. Avila Pulgarín, Leidy Steffany, Acevedo Restrepo, Andrés, Jurado Guevara, Jairo Andrés, Polanco Echeverry, Diana, Velásquez Vélez, Raúl, & Zapata Salas, Richard. (2013). Infección por hemoparásitos en caprinos y ovinos de apriscos de cinco municipios del norte y nororiente de Antioquia (Colombia). **CES Medicina Veterinaria y Zootecnia**, 8(1), 11-21.
7. B. Adehanom, D. Dagnachew, Teshale Teklue and N. Surendra, 2015. Validation of the FAMACHA© Eye Color Technique for Detecting Anemic Sheep and Goats in Jigjiga Zone of Somali Región, Eastern Ethiopia. **Research Journal of Veterinary Sciences**, 8: 61-67
8. Baneth G. (2014). Tick-borne infections of animals and humans: a common ground. **Int J Parasitol**, 44(9):591-6. doi: 10.1016/j.ijpara.2014.03.011. Epub 2014 May 15. PMID: 24846527.
9. Bauer, B. U., Răileanu, C., Tauchmann, O., Fischer, S., Ambros, C., Silaghi, C., & Ganter, M. (2021). *Anaplasma phagocytophilum* and *Anaplasma ovis*—emerging pathogens in the German sheep population. **Pathogens**,10(10), 1298.
10. Calderón, A., Martínez, N., & Iguarán, H. (2016). Frecuencia de hematozoarios en bovinos de una región del caribe colombiano. **Revista UDCA Actualidad & Divulgación Científica**, 19(1), 131-138.
11. Canul, A. J. C., Pérez-Hernández, R., Salazar-Cuytun, R., García-Herrera, R. A., Herrera-Camacho, J., & López-Duran, S. K. (2022). Condición corporal en ovejas Pelibuey en el trópico de México. **Boletín de Ciencias Agropecuarias del ICAP**, 8(16), 31-35.
12. Chochlakis, D., Ioannou, I., Sharif, L., Kokkini, S., Hristophi, N., Dimitriou, T., & Psaroulaki, A. (2009). Prevalence of *Anaplasma* sp. in goats and sheep in Cyprus. **Vector-Borne and Zoonotic Diseases**, 9(5), 457-463.
13. Coello-Peralta, R. D., Solórzano, N. C. C., Mazamba, M. D. L. S., Echeverria, E. O. R., Velásquez, L. L. A., & Gómez, B. J. P. (2021). Natural infection of *Trypanosoma* sp. in domestic sheep from Ecuador. **Ciência Rural**, 52, e20210141.
14. De La Fuente J., Atkinson M. W., Naranjo V., Fernández De Mera I. G., Mangold A. J., Keating K. A., and Kocan K. M. (2007). Sequence analysis of the *msp4* gene of *Anaplasma ovis* strains, **Veterinary Microbiology**, 119 (2–4): 375–381. <https://doi.org/10.1016/j.vetmic.2006.09.011>
15. Di Loria, A., Veneziano, V., Piantedosi, D., Rinaldi, L., Cortese, L., Mezzino, L., Ciaramella, P. (2009). Evaluación del sistema FAMACHA para detectar la gravedad de la anemia en ovejas del sur de Italia, **Parasitología veterinaria**, 161(1-2), 53–59.
16. Dill, D. B., & Costill, D. L. (1974). Calculation of Percentage in Changes in Volumes of Blood, Plasma, and Red Cells in Dehydration. **Journal of Applied Physiology**, 37, 247-248.

17. Egbe-Nwiyi TN, Sherrif GA, Paul BT, 2018. Prevalence of tick-borne haemoparasitic diseases (TBHDS) and haematological changes in sheep and goats in Maiduguri abattoir. **J. Vet. Med. Anim. Health**, 10: 28-33.
18. Eisawi, N. M., El Hussein, A. R. M., Hassan, D. A., Musa, A. B., Hussien, M. O., Enan, K. A., & Bakheit, M. A. (2020). A molecular prevalence survey on Anaplasma infection among domestic ruminants in Khartoum State, Sudan. **Tropical Animal Health and Production**, 52, 1845-1852
19. Flórez M, Julio, Hernández P, Marcos, Bustamante Y, Moris, & Vergara G, Oscar. (2020). Caracterización morfo estructural e índices zoométricos de hembras Ovino de Pelo Criollo Colombiano "OPC" Sudán. **Revista MVZ Córdoba**, 25(3), 116-125.
20. González, B. C., Obregón, D., Alemán, Y., Alfonso, P., Vega, E., Díaz, A., & Martínez, S. (2014). Tendencias en el diagnóstico de la anaplasmosis bovina. **Revista de Salud Animal**, 36(2), 73-79.
21. Hamzah, K. J., & Hasso, S. A. (2019). Molecular prevalence of Anaplasma phagocytophilum in sheep from Iraq. **Open Veterinary Journal**, 9(3), 238-245.
22. Hernández D, Montes D, De la Ossa J. (2020). Asociación del polimorfismo FecB con la prolificidad natural del Ovino de Pelo Colombiano. **Rev MVZ Córdoba**, 25(1):1–6. <https://doi.org/10.21897/rmvz.17715>.
23. Instituto Colombiano Agropecuario (ICA)-Republica de Colombia. (2023). Censo pecuario año 2023. <https://www.ica.gov.co/areas/pecuaria/servicios/epidemiologia-veterinaria/censos-2016/censo-2018>
24. Jaimes-Duñez, J., Triana-Chávez, O., & Mejía-Jaramillo, A.M. (2017). Parasitological and molecular surveys reveal high rates of infection with vector-borne pathogens and clinical anemia signs associated with infection in cattle from two important livestock areas in Colombia. **Ticks and Tick-Borne Diseases**, 8(2), 290-299.
25. Li, Y., Galon, E. M., Guo, Q., Rizk, M. A., Moumouni, P. F. A., Liu, M., & Xuan, X. (2020). Molecular detection and identification of Babesia spp., Theileria spp., and Anaplasma spp. in sheep from border regions, northwestern China. **Frontiers in Veterinary Science**, 7, 630.
26. López-Jácome, L. E., Hernández-Durán, M., Colín-Castro, C. A., Ortega-Peña, S., Cerón-González, G., & Franco-Cendejas, R. (2014). Las tinciones básicas en el laboratorio de microbiología. **Investigación en discapacidad**, 3(1): 10-18.
27. Manterola, C., Otzen, T. (2014). Estudios observacionales. Los diseños utilizados con mayor frecuencia en investigación clínica. **Int. J. Morphol**, 32(2):634-645.
28. Martínez J., Tatis F. Incidencia de los hemoparásitos en la producción ovina en condiciones de pastoreo extensivo en el municipio de Toluviejo, Sucre. Tesis de pregrado, Facultad de Ciencias Agropecuarias, Universidad de Sucre, Sincelejo, 2001. 67 p.
29. Montes-Vergara, D., Hernández-Herrera, D., & Carrillo-González, D. (2022). Caracterización morfológica, faneróptica y de genes dominantes de la gallina criolla Subespecie nudicollis en la región Sabanas del departamento de Sucre-Colombia. **Revista MVZ Córdoba**, 27(1), 2022. <https://doi.org/10.21897/rmvz.2599>
30. Naeem, M., Amaro-Estrada, I., Taqadus, A., Swelum, A. A., Alqhtani, A. H., Asif, M., & Iqbal, F. (2023). Molecular prevalence and associated risk factors of Anaplasma ovis in Pakistani sheep. **Frontiers in Veterinary Science**, 10, 1096418.
31. Nangru, A., Maharana, BR, Vohra, S., Kumar, B., Ganguly, A., Sahu, S., Khichar, V. (2023). Identification, molecular characterization and risk factors of Theileria infection among sheep: a first comprehensive report from North India. **Animal Biotechnology**, 34 (8), 3658–3670. <https://doi.org/10.1080/10495398.2023.2189928>
32. Noaman, V., Sazmand, A. Anaplasma ovis infection in sheep from Iran: molecular prevalence, associated risk factors, and spatial clustering. **Trop Anim Health Prod**, 54(1): 1-6 (2022). <https://doi.org/10.1007/s11250-021-03007-4>

33. Noriega-Márquez, J., Hernández-Herrera, D., Bustamante-Yáñez, M., Álvarez-Franco, L., Ariza-Botero, M., Palacios-Erazo, Y., & Vergara-Garay, O. (2022). Curvas de crecimiento en ovino de pelo colombiano en los departamentos de Córdoba y Cesar, Colombia. **Revista UDCA Actualidad & Divulgación Científica**, 25(2): 1-8. <https://revistas.udca.edu.co/index.php/ruadc/article/view/1727/2455>
34. Nyifi, A. S., & Bilbonga, G. (2023). Effect of Tick-Borne Haemoparasitic Diseases on Haematological Parameters of Small Ruminants Managed Under Semi-intensive System in Wukari Town Taraba State Nigeria. **International Journal of Research and Scientific Innovation**, 10(11), 457-465.
35. Nyifi, A., Ahmadu, P., & Mamtso, R. (2023). Prevalence of anaplasmosis in small ruminants in some selected livestock farms in Wukari metropolis taraba State Nigeria. **FUDMA Journal of Agriculture and Agricultural Technology**, 9(3), 52-57.
36. Onyiche, T. E., & MacLeod, E. T. (2023). Hard ticks (Acari: Ixodidae) and tick-borne diseases of sheep and goats in África: A review. **Ticks and tick-borne diseases**, 14(6), 102232.
37. Onyiche, T. E., Mofokeng, L. S., Thekiso, O., & MacLeod, E. T. (2022). Molecular survey for tick-borne pathogens and associated risk factors in sheep and goats in Kano Metropolis, Nigeria. **Veterinary Parasitology: Regional Studies and Reports**, 33, 100753.
38. Plaza Cuadrado, Alberto., Hernández Padilla, Eduardo., Rugeles Pinto, Clara., Vergara
39. Garay, Oscar., Herrera Benavides, Yonairo (2019). Perfil hematológico durante la gestación de Ovinos de Pelo Criollos en el departamento de Córdoba, Colombia. **Revista Colombiana de Ciencia Animal**, 11(1). <https://revistas.unisucre.edu.co/index.php/recia/article/view/657/765>
40. Prajapati, A., Prajapati, B., Patel, A., Chauhan, P., Das, B., Raval, S & Patel, R. (2023). Molecular identification and genetic characterization of Theileria and Anaplasma infection in sheep and goat of North Gujarat, India. **Parasitology Research**, 122(6), 1427-1433.
41. R Core Team (2023). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.
42. Rahravani, M., Moravedji, M., Mostafavi, E., Mozoun, M. M., Zeeyaie, A. H., Mohammadi, M., & Ameri, M. (2023). Clinical, hematological and molecular evaluation of piroplasma and Anaplasma infections in small ruminants and tick vectors from Kurdistan province, western Iran. **Research in Veterinary Science**, 159, 44-56.
43. Reategui-Valles, M. O., & López-Flores, A. M. (2023). Prevalencia de Babesiosis bovina en el distrito de Cuñumbuqui, Perú. **Revista De Veterinaria Y Zootecnia Amazónica**, 3(2). e560. <https://doi.org/10.51252/revza.v3i2.560>
44. Russel AJF, Doney JM, Gunn RG (1969) Subjective assessment of body fat in live sheep. **J Agric Sci**, 72(3):451-4
45. Şahin, Ö., Aytakin, İ., Boztepe, S. et al. (2021). Relationships between FAMACHA© scores and parasite incidence in sheep and goats. **Trop Anim Health Prod**, 53(2): 331. <https://doi.org/10.1007/s11250-021-02769-1>
46. Shah, S. S. A., Khan, M. I., & Rahman, H. U. (2017). Epidemiological and hematological investigations of tick-borne diseases in small ruminants in Peshawar and Khyber Agency, Pakistan. **J. adv. parasitol**, 4(1), 15-22.
47. Tamrat, H., Tagel, W., & Belayneh, N. (2023). Epidemiology of Ixodid tick infestation and tick-borne haemopathogens in small ruminant from Enarje Enawuga, North Western Ethiopia. **Veterinary Medicine and Science**, 9(3), 1318-1326.
48. Taqadus, A., Chiou, C. C., Amaro-Estrada, I., Asif, M., Nasreen, N., Ahmad, G., & Chen, C. C. (2023). Epidemiology and phylogeny of Anaplasma ovis with a note on hematological and biochemical changes in asymptomatic goats enrolled from four districts in Punjab, Pakistan. **Vector-Borne and Zoonotic Diseases**, 23(10), 495-506.
49. Torres Rodríguez, A., Pérez, N. I., Cruz, A. A., & Záldivar, Y. L. (2023). Relación entre los valores hematológicos del hemograma y la carga parasitaria presente en ovinos Pelibuey resilientes en condiciones de pastoreo. **Revista de Producción Animal**, 35(3), 5-21.

50. Torres Torres, A. A., Lara Díaz, M. I., & Páez Díaz, R. (2021). Factores que influyen en la presentación actual de *Anaplasma* sp. y *Babesia* spp. en bovinos en el trópico. **Biociencias (UNAD)**, 5(1): 155-181.
51. Valente, J. D., Mongruel, A. C., Machado, C. A., Chiyo, L., Leandro, A. S., Britto, A. S., & Vieira, R. F. (2019). Tick-borne pathogens in carthorses from Foz do Iguacu City, Paraná State, southern Brazil: A tri-border area of Brazil, Paraguay and Argentina. **Veterinary parasitology**, 273: 71-79.
52. Vargas-Cuy, D. H., Torres-Caycedo, M. I., & Pulido-Medellín, M. O. (2019). Anaplasmosis y babesiosis: estudio actual. *Pensamiento y Acción*, (26): 45–60. Recuperado a partir de [https://revistas.uptc.edu.co/index.php/pensamiento\\_accion/article/view/9723](https://revistas.uptc.edu.co/index.php/pensamiento_accion/article/view/9723)
53. Woo P. (1971). Evaluation of the haematocrit centrifuge and other techniques for the field diagnosis of human tripanosomiasis and filariasis. **Acta Trop**, 28(3):298-303.